



Review

# Long COVID and the Neuroendocrinology of Microbial Translocation Outside the GI Tract: Some Treatment Strategies

Adonis Sfera <sup>1,2,\*</sup>, Carolina Osorio <sup>3</sup>, Sabine Hazan <sup>4</sup>, Zisis Kozlakidis <sup>5</sup>, Jose Campo Maldonado <sup>6</sup>, Carlos Manuel Zapata-Martín del Campo <sup>7</sup>, Jonathan J. Anton <sup>8</sup>, Leah Rahman <sup>9</sup>, Christina V. Andronescu <sup>10</sup> and Garth L. Nicolson <sup>11</sup>

<sup>1</sup> Patton State Hospital, San Bernardino, CA 92407, USA

<sup>2</sup> Department of Psychiatry, University of California, Riverside, CA 92521, USA

<sup>3</sup> Department of Psychiatry, Loma Linda University, Loma Linda, CA 92354, USA

<sup>4</sup> ProgenomaBiome, Ventura, CA 93003, USA

<sup>5</sup> International Agency for Research on Cancer World Health Organization, 69372 Lyon, France

<sup>6</sup> Department of Medicine, The University of Texas Rio Grande Valley, Edinburg, TX 78539, USA

<sup>7</sup> Instituto Nacional de Cardiología and in Ignacio Chávez, Mexico City 14080, Mexico

<sup>8</sup> Department of Biology, California Baptist University, Riverside, CA 92521, USA

<sup>9</sup> Department of Neuroscience, University of Oregon, Eugene, OR 97402, USA

<sup>10</sup> Medical Anthropology Department, Stanford University, Stanford, CA 94305, USA

<sup>11</sup> Department of Molecular Pathology, The Institute for Molecular Medicine, Huntington Beach, CA 92647, USA

\* Correspondence: adonis.sfera@dsh.ca.gov



**Citation:** Sfera, A.; Osorio, C.; Hazan, S.; Kozlakidis, Z.; Maldonado, J.C.; Zapata-Martín del Campo, C.M.; Anton, J.J.; Rahman, L.; Andronescu, C.V.; Nicolson, G.L. Long COVID and the Neuroendocrinology of Microbial Translocation Outside the GI Tract: Some Treatment Strategies. *Endocrines* **2022**, *3*, 703–725. <https://doi.org/10.3390/endocrines3040058>

Academic Editors: Giuseppe Lisco and Vincenzo Triggiani

Received: 30 September 2022

Accepted: 25 October 2022

Published: 7 November 2022

**Publisher's Note:** MDPI stays neutral with regard to jurisdictional claims in published maps and institutional affiliations.



**Copyright:** © 2022 by the authors. Licensee MDPI, Basel, Switzerland. This article is an open access article distributed under the terms and conditions of the Creative Commons Attribution (CC BY) license (<https://creativecommons.org/licenses/by/4.0/>).

**Abstract:** Similar to previous pandemics, COVID-19 has been succeeded by well-documented post-infectious sequelae, including chronic fatigue, cough, shortness of breath, myalgia, and concentration difficulties, which may last 5 to 12 weeks or longer after the acute phase of illness. Both the psychological stress of SARS-CoV-2 infection and being diagnosed with COVID-19 can upregulate cortisol, a stress hormone that disrupts the efferocytosis effectors, macrophages, and natural killer cells, leading to the excessive accumulation of senescent cells and disruption of biological barriers. This has been well-established in cancer patients who often experience unrelenting fatigue as well as gut and blood-brain barrier dysfunction upon treatment with senescence-inducing radiation or chemotherapy. In our previous research from 2020 and 2021, we linked COVID-19 to myalgic encephalomyelitis/chronic fatigue syndrome (ME/CFS) via angiotensin II upregulation, premature endothelial senescence, intestinal barrier dysfunction, and microbial translocation from the gastrointestinal tract into the systemic circulation. In 2021 and 2022, these hypotheses were validated and SARS-CoV-2-induced cellular senescence as well as microbial translocation were documented in both acute SARS-CoV-2 infection, long COVID, and ME/CFS, connecting intestinal barrier dysfunction to disabling fatigue and specific infectious events. The purpose of this narrative review is to summarize what is currently known about host immune responses to translocated gut microbes and how these responses relate to fatiguing illnesses, including long COVID. To accomplish this goal, we examine the role of intestinal and blood-brain barriers in long COVID and other illnesses typified by chronic fatigue, with a special emphasis on commensal microbes functioning as viral reservoirs. Furthermore, we discuss the role of SARS-CoV-2/Mycoplasma coinfection in dysfunctional efferocytosis, emphasizing some potential novel treatment strategies, including the use of senotherapeutic drugs, HMGB1 inhibitors, Toll-like receptor 4 (TLR4) blockers, and membrane lipid replacement.

**Keywords:** cortisol; HMGB1; microbial translocation; SARS-CoV-2

## 1. Introduction

In the post-pandemic era, residual or long COVID-19 sequelae have been gradually emerging as many patients experience prolonged fatigue, cough, shortness of breath, myalgia, and problems with concentration long after the acute illness phase [1]. From a biological

pathway perspective, as both SARS-CoV-2 infection and the associated psychological stress upregulate cortisol, the function of macrophages and natural killer (NK) cells may be impaired, disrupting the clearance of senescent, damaged, or virus-infected cells. This may lead to biological barrier dysfunction and chronic fatigue, phenomena well-documented in cancer survivors treated with cellular senescence-inducing chemotherapy or radiation [2–5].

At the molecular level, upregulated cortisol lowers the expression of *claudin-1* (CLDN1), an intestinal tight junction protein, facilitating microbial translocation outside of the gastrointestinal (GI) tract [6]. In the central nervous system (CNS), *P-glycoprotein* (P-gp), a cortisol substrate, functions as a gatekeeper of the blood–brain barrier (BBB), likely accounting for hypercortisolemia-increased barrier permeability [7–9]. Interestingly, in the GI tract, gut microbiota and lipopolysaccharide (LPS), a cell wall component of Gram-negative bacteria, regulate P-gp, connecting this protein to microbial translocation [10,11].

Recent studies have found a direct relationship between circulating levels of cortisol and premature cellular senescence, a phenotype characterized by permanent cell cycle arrest, active metabolism, and a detrimental secretome [12]. The accumulation of senescent cells was found to be associated not only with organismal aging but also with chronic fatigue, pain, and depression, documented in cancer survivors [2]. On the other hand, enhanced elimination of senescent cells via senotherapeutics can correct barrier dysfunction, lowering fatigue [13–15].

NK cells and macrophages can execute the phagocytic engulfment (efferocytosis) of damaged or dead cells, including malignant, virus-infected, and senescent cells. Dysfunctional efferocytosis has been associated with biological barrier disruption, inflammatory bowel disease (IBD), and a constellation of symptoms reminiscent of long COVID and other fatiguing illnesses [16–21]. Indeed, NK cell dysfunction is one of the most consistent findings in long COVID-19, myalgic encephalomyelitis/chronic fatigue syndrome (ME/CFS), Gulf War illnesses (GWI), and fibromyalgia (FM), linking these pathologies to dysfunctional efferocytosis [22–27]. Moreover, NK cells had been previously linked to fatigue-associated thyroid, adrenal, hypothalamic, and pituitary disorders, thus connecting these neuroendocrine-related pathologies with dysfunctional immunity [28–32]. Furthermore, NK cells express estrogen, prolactin, and cortisol receptors as well as a functional renin-angiotensin-system (RAS), rendering them sensitive to hormonal fluctuations [33–35]. NK cells are capable of paracrine signaling and they secrete biomolecules, including *perforin*, *granzyme B*, and the *high mobility group box 1 protein* (HMGB1) that can facilitate the elimination of damaged, senescent, and/or malignant cells [36–38].

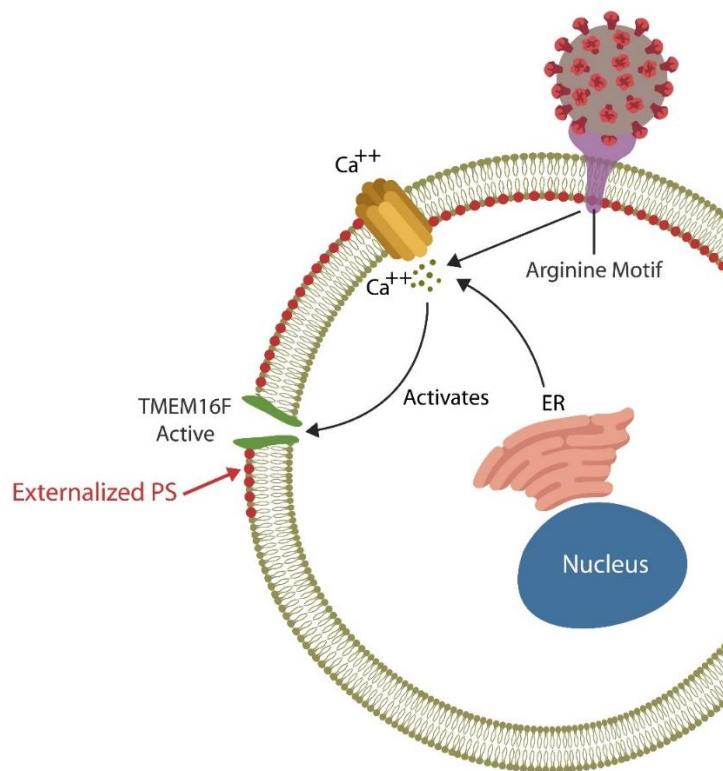
Microbial translocation markers, such as LPS, lipopolysaccharide binding protein (LPB), soluble CD14 (sCD14), and HMGB1, were found to be elevated in long COVID, highlighting the role of dysfunctional gut barrier in this condition [30–41]. Indeed, accumulation of senescent cells has been associated with HMGB1 spillover into the extracellular space where it can act as an inflammagen and barrier disruptor [42–46]. Therefore, up-regulated HMGB1, documented in ME/CFS, FM, and GWI, and COVID-19, directly links dysfunctional efferocytosis to chronic fatigue [47–50]. For example, gut HMGB1 disrupts the barrier tight junctions and is considered a biomarker of inflammatory bowel disease (IBD), a condition associated with both fatigue and increased GI tract permeability. This may explain certain SARS-CoV-2 bacteriophage-like properties as this virus can likely penetrate the microbial cell walls [51–57].

In this article, we will take a closer look at the role of dysfunctional NK cells and efferocytosis in long COVID and other fatiguing illnesses. We also discuss the comorbidity of *Mycoplasma* species and SARS-CoV-2 as well as their coinfection, barrier rehabilitation via senotherapeutic drugs, HMGB1 inhibitors, Toll-like receptor 4 (TLR-4) blockers and membrane lipid replacement (MLR).

## 2. Efferocytosis and Biological Barriers

Each day billions of cells throughout the body undergo apoptosis and are removed by professional phagocytes, macrophages, monocytes, and neutrophils as well as non-

professional phagocytes, including the intestinal epithelial cells (IECs) and the M2 microglia in the BBB [58–60]. Professional and non-professional phagocytes are assisted by NK cells that can eliminate defective and pathogen-infected cells without prior sensitization [61–63]. NK cells maintain the integrity of BBB and the intestinal barrier as they can promptly clear damaged cells, preventing inflammation and barrier disruption [64,65]. To accomplish this, NK cells perforate the membrane of targeted cells by releasing HMGB1, perforin, and granzyme, triggering apoptosis by  $\text{Ca}^{2+}$  influx [66,67]. Upregulated cytosolic  $\text{Ca}^{2+}$  is known to activate *TMEM16F*, an enzyme that flips phosphatidylserine (PS) to the outer leaflet of the cell membrane, providing a distress signal, that attracts immune cells, promoting phagocytosis [68]. Exposed PS (ePS) comprises an “eat me” or “fuse with me” signal that can lead to either cell death or syncytia formation, depending on the degree of cell membrane damage [69]. For example, less damaged cells can fuse with each other for protection, a phenomenon documented in many tissues, including the CNS [70] (Figure 1). Several studies have demonstrated that senescent and cancer cells can avoid elimination by expressing CD47, a “don’t eat me” signal, that inhibits phagocytosis [71–73].



**Figure 1.** The SARS-CoV-2 receptor binding site (RBS) contains a double arginine insert (PRRA) or arginine motif, that perforates the cell membrane, triggering  $\text{Ca}^{2+}$  influx from both the endoplasmic reticulum (ER) and the extracellular compartment. Upregulated cytosolic  $\text{Ca}^{2+}$  activates TMEN16F, externalizing phosphatidylserine (ePS), an “eat me” or “fuse me” signal that leads to cell death (if the damage is irreparable) or cell–cell fusion (if the cell can be repaired). Cell–cell fusion induces premature cellular senescence, disrupting biological barriers. The virus benefits from ePS as this comprises a global immunosuppressive signal, allowing its undetected entry into host cells.

### 2.1. Blood–Brain Barrier

The BBB, a highly regulated interface between the circulatory system and the CNS, consists of cerebral endothelial cells (ECs) that regulate the inward and outward movement of molecules and ions into the CNS [74]. BBB disruption enables viral entry into the brain along with inflammatory cells, and/or deleterious molecules that can trigger infection. Indeed, members of at least 11 viral families, including the human immunodeficiency

virus-1 (HIV-1), T-cell leukemia virus, lymphocytic choriomeningitis virus, West Nile virus, and others, can enter the brain, causing encephalitis [75–77].

COVID-19-mediated accumulation of senescent ECs compromises the BBB, allowing viral and microbial access to the CNS [78]. In contrast, enhanced elimination of senescent cells via senolytic drugs decreases COVID-19 mortality in rodents, highlighting the role of senescent cells in BBB dysfunction [79,80]. In addition, the S protein of the SARS-CoV-2 virus was demonstrated to directly bind bacterial LPS, outlining a virus-mediated mechanism of endotoxin entry into the CNS [40–81]. LPS-induced neuroinflammation has been associated with microglial fusion and multinucleation, generating highly phagocytic phenotypes that can cause collateral damage by eliminating viable neurons [82,83]. Indeed, long COVID has been associated with LPS-activated microglia (M1 phenotype), neuroinflammation and neuronal death [82–85]. On the other hand, the M2 microglial phenotype has been shown to repair the damage, protecting the neurons [58].

## 2.2. Intestinal Barrier

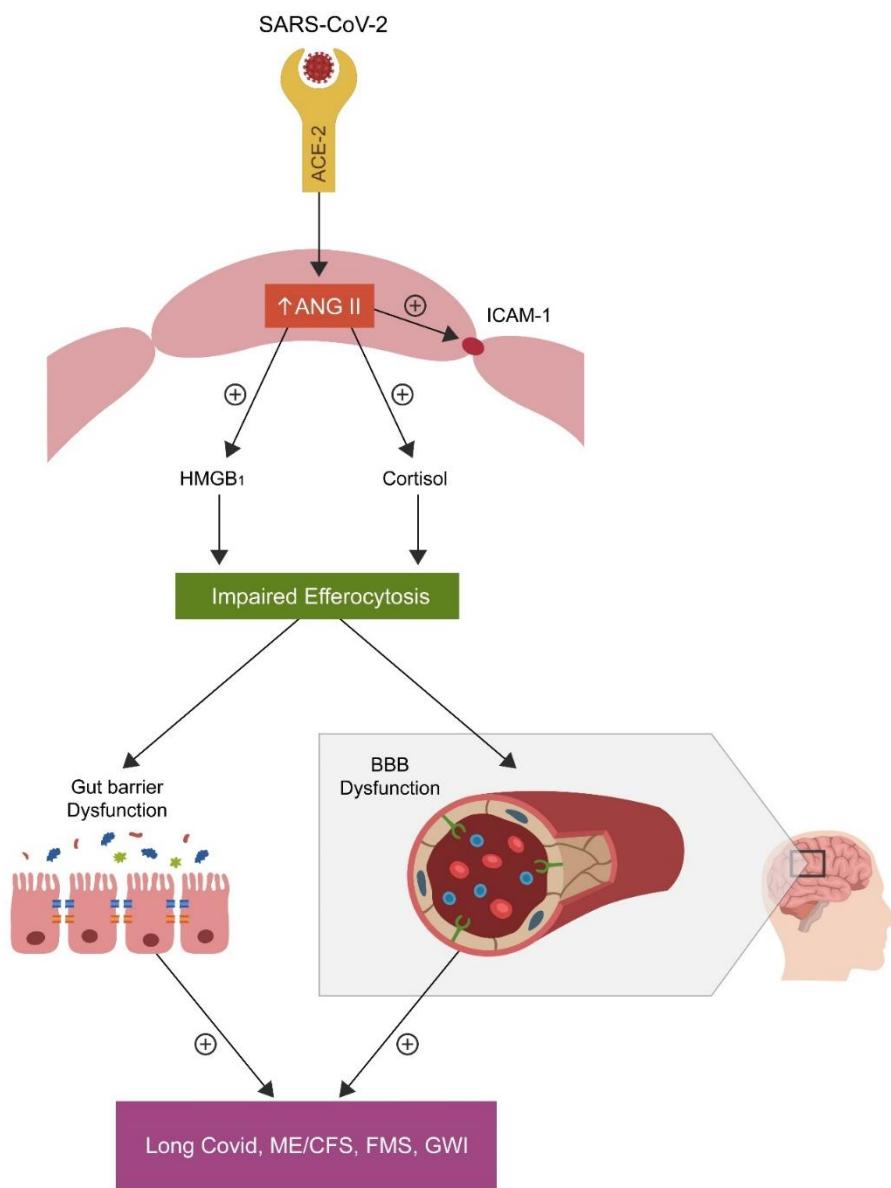
Many viruses, including SARS-CoV-2, enhance infectivity by usurping both the physiological cell–cell fusion and efferocytosis, disrupting biological barriers [75,86]. For example, the SARS-CoV-2 virus thrives in infected cells and likely inhibits their clearance, causing inflammation and barrier dysfunction [87]. In addition, SARS-CoV-2 promotes pathological cell–cell fusion and syncytia formation by generating cell membrane pores via the PRRA (proline-arginine-arginine-alanine) motif situated at the furin-cleavage site (FCS). This system is reminiscent of microbial twin arginine translocation pathway, a pore-forming mechanism implicated in bacterial virulence [88–90]. Moreover, SARS-CoV-2 fusion with Mycoplasma, an arginine dependent microorganism, may explain the high comorbidity of these very different infections [91]. Cell membrane pores lead to ePS, a global immunosuppressive signal, that helps the virus exploit host defenses [92]. The subsequent, syncytia formation can then induce premature cellular senescence and the release of senescence-associated secretory phenotype (SASP), a pathological secretome that disrupts endothelial barriers by promoting premature ECs senescence, a phenotype documented in both ME/CFS and COVID-19 [42,93–95].

Senescence-induced pathological syncytia can trigger lymphopenia by cell-in-cell phenomena, elimination of viable lymphocytes, including NK cells, a frequent finding in ME/CFS [96–98]. In addition, as cellular senescence upregulates HMGB1, it may further predispose to fatiguing disorders [44,47–49,99–102].

In the GI tract, IECs comprise a single layer of tightly linked columnar cells that are short-lived and need to be replaced every 4 to 5 days to maintain an adequate barrier function [103]. Moreover, IECs acting as non-professional phagocytes, can engulf the translocating microbes and/or antigens, preventing microbial translocation outside the GI tract [104]. However, accumulation of uncleared, defective IECs can trigger inflammation, predisposing to IBD and other illnesses marked by dysfunctional barrier [105]. Macrophages, intestinal NK cells, and Paneth cells contribute to barrier integrity by promptly removing damaged IECs, thus averting necrosis and inflammation-mediated pathology [21,106–108]. Interestingly, IECs were demonstrated to produce cortisol, a steroid hormone that lowers the expression of CLDN1 that in return increases intestinal permeability [109–111].

The role of ANG II: COVID-19-upregulated *angiotensin II* (ANG II) can disrupt efferocytosis inducing ECs senescence and vascular barrier dysfunction via *angiotensin II type 1 receptors* (AT-1Rs) which can enhance both cortisol and HMGB1 production, [112,113] (Figure 2).

Increased cytosolic ANG II disrupts the function of NK cells and, as these cells express a viable RAS, including ACE-2, the virus may directly infect these immune cells [114,115]. In the CNS, ANG II disrupts the BBB via AT1 receptors; in contrast, Losartan, an AT1 antagonist, can repair this and the intestinal biological barrier [116,117].



**Figure 2.** SARS-CoV-2 attachment to ACE-2, blocks this enzyme, causing angiotensin II (ANG II) accumulation by inhibiting its hydrolysis. Upregulated ANG II, increases both cortisol and HMGB1, disrupting the efferocytosis of senescent cells. Accumulation of senescent cells triggers inflammation and biological barrier disruption, a common pathology found not only in the disorders marked by chronic fatigue but also in neuropsychiatric and autoimmune diseases.

### 3. Biological Barriers and Chronic Fatigue

In previous research (2020 and 2021), we linked COVID-19 to ME/CFS via ANG II, endothelial senescence, intestinal barrier dysfunction, and microbial translocation from the GI tract into host systemic circulation [118,119]. Subsequently, these notions were validated by various groups and virus-induced cellular senescence as well as microbial translocation were documented in both acute COVID-19 illness and long COVID, linking intestinal barrier dysfunction to chronic fatigue [120–127]. Indeed, abnormal intestinal permeability has been documented in GWI, COVID-19 and FM, connecting these conditions to microbial translocation [127–129]. Aside from fatiguing illnesses, increased microbial migration outside the GI tract has also been demonstrated in autoimmune disorders, cancers, and neuropsychiatric conditions, including schizophrenia and Alzheimer’s disease (AD) [130–134].

Numerous gut microbes express proteins that are identical or similar to those of the human host, eliciting antibodies upon translocation that could be misconstrued as autoantibodies. For example, several intestinal microbial species express *QseC*, an adrenergic receptor, indicating that antibodies against translocated *QseC* could trigger human pathology [135]. Indeed, autoantibodies against  $\beta 1$  adrenergic receptors were found in patients with ME/CFS as well as in certain cardiovascular diseases, linking these conditions to microbial translocation [136,137]. In addition, bacteria and archaea synthesize acetylcholine (ACh) and express nicotinic or muscarinic receptors that can elicit antibodies upon translocation, blurring the distinction between autoimmunity and immune reactions to gut microbes at extra-intestinal locations [138]. For example, antimuscarinic autoantibodies, documented in ME/CFS, may represent antibodies against translocating microbes expressing ACh receptors [139]. Indeed, ACh receptor autoantibodies, markers of myasthenia gravis and Lambert–Eaton syndrome, have also been documented in *Mycoplasma pneumoniae* and influenza, suggesting that, rather than autoantibodies, these could be conventional antibodies against translocated pathogens [140–144]. *Mycoplasma* infection has also been associated with autoantibodies against *N-methyl-D-aspartate* (NMDA) receptors, markers of neuropsychiatric disorders, including schizophrenia, connecting this illness to gut microbes [145,146]. Furthermore, microbiota-expressing *metabotropic glutamate receptors subtype 2* (mGluR2), linked to both SARS-CoV-2 and Rabies (RABV) viruses, may elicit antibodies rather than autoantibodies, further blurring the border between immunity and autoimmunity [147,148]. For example, mGluR2 autoantibodies, documented in paraneoplastic syndrome as well as in *Mycoplasma* and Herpes virus infection, may be conventional immunoglobulins against translocated microbes [149–151]. Together this data raises an important question: are antibodies against translocated microbial proteins being misidentified as autoantibodies [152,153]?

The human gut microbiome is composed of bacteria, yeasts, fungi, and viruses, an ecosystem in which microbes can inhibit some viral pathogens, while promoting others [154,155]. Most gut viruses are bacteriophages (bacteria-infecting viruses); however, others including SARS-CoV-2, may display bacteriophage-like properties and enter selective microbes, including the cell wall-deficient *Mycoplasmas* [156–158]. In addition, angiotensin converting enzyme-2 (ACE-2), the SARS-CoV-2 entry portal, is expressed by some gut microbes, indicating that the SARS-CoV-2 virus may enter some microbiota, potentially utilizing them as reservoirs [22]. Indeed, long COVID was found to reactivate Epstein–Barr virus (EBV), suggesting that translocating microbes, containing SARS-CoV-2, could play a key role in maintaining a state of latent infection [159].

When thinking about this problem, studying the microbiome may allow us to reconceptualize some autoimmune disorders as conventional immunity against translocating microbes or their antigens, a model that has already been proposed in the etiopathogenesis of systemic lupus erythematosus (SLE) [160,161]. This is significant as methotrexate, a drug often prescribed to patients with autoimmune disorders, can increase intestinal permeability, further facilitating microbial translocation, while at the same time, lowering host immune defenses that oppose these agents [162–164].

#### 4. Rethinking *Mycoplasma*

*Mycoplasmas*, members of the Mollicutes class of bacteria, are among the simplest and smallest known self-replicating microorganisms [165]. Their genomes, containing about 400–600 genes, are comprised of either a single stranded RNA or a double stranded DNA nucleic acid. *Mycoplasmas* can be commensal or pathogenic, the former dwelling superficially, for example in the oral cavity, while the latter inside host cells [166,167]. Like SARS-CoV-2, some *Mycoplasma* species release immune modulators and proinflammatory cytokines that can disrupt host immunity or cause hyperinflammatory reactions (“cytokine storms”) [168–171]. In addition, some *Mycoplasma* species synthesize arginine deaminase (ADI), an enzyme that can further disrupt host immunity [172,173].

The infection with SARS-CoV-2 virus results in variable patient outcomes, ranging from few or no symptoms, in some individuals, to critical illness and death in a small number of patients. As viral infections are often accompanied by secondary bacterial contagion, coinfection may contribute to the majority of unfavorable outcomes. Indeed, *Mycoplasma* comorbidity has been associated with poor COVID-19 prognosis, while epidemiological studies show up to 47% comorbidity between SARS-CoV-2 and *Mycoplasma* [174–180]. In addition, like SARS-CoV-2, *Mycoplasma* infections have been associated with BBB dysfunction and IBD, connecting this cell wall-deficient bacterium to biological barrier dysfunction [181–183].

Coinfection with *Mycoplasma species* has been demonstrated in other viral illnesses, including HIV-1, HHV-6, and various influenza strains as well as in some fatiguing illnesses and cancers. This suggests that *Mycoplasma* may thrive in defective cells and probably induce further cellular damage by disrupting efferocytosis [184–187]. There are also significant overlaps in the clinical picture, laboratory, and imaging studies between SARS-CoV-2 and *Mycoplasma* infections, further complicating the differential diagnosis [188–190]. Moreover, diagnostic tests, including *Mycoplasma species* serology, culture, and even nucleic acid amplification, such as PCR, are marked by numerous limitations [191–193]. In this regard, false-positive and -negative COVID-19 serological test results have been reported in many patients with *Mycoplasma pneumoniae* infection, highlighting the limitation of these assays [194,195]. The next generation sequencing by shot gun methodology appears promising for differentiating *Mycoplasma* from SARS-CoV-2 and may have a place in the diagnosis of long COVID [196]. However, leukopenia, lymphocytopenia, thrombocytopenia, and thromboembolism were documented in both SARS-CoV-2 and *Mycoplasma* infections, further highlighting their intertwined etiopathogenesis [197–200]. Furthermore, certain anti-microbial treatments, such as azithromycin or tetracyclines, were found beneficial for both *Mycoplasma* and COVID-19, further suggesting a likely silent partnership between these quite different infections [201,202].

Several *Mycoplasma* species express the integrin motif, Arg-Gly-Asp, or RGD, a cell attachment sequence that connects these pathogens to the host extracellular matrix (ECM) proteins, including *integrins*, *laminins*, and *fibronectin* (FBN) [169,203–206]. A recent milestone in COVID-19 pathogenesis was the revelation that SARS-CoV-2 receptor binding site (RBS) contains an RGD motif that could facilitate viral entry in host cells [207–209]. Since both SARS-CoV-2 and *Mycoplasma species* bind FBN and express the RGD motif, they may fuse with each other, engendering a combined pathology [206,209,210]. In addition, it has been established that *Mycoplasma fermentans incognitus* strain stimulates tissue plasminogen activator (tPA) which converts plasminogen to plasmin, a protein that, like furin, can cleave the SARS-CoV-2 S antigen at the S1/S2 site, triggering pathological cell-cell fusion [211–213]. Elevated plasmin and plasminogen levels are common findings in severe COVID-19 illness as well as in patients with various chronic diseases, including hypertension, diabetes, and cardiovascular diseases. This may account for an unfavorable COVID-19 prognosis in patients with these disorders [214,215]. Moreover, SARS-CoV-2 may benefit from its association with *Mycoplasma* as this bacterium can directly block host immunoglobulins, protecting the virus [216,217]. This finding may be significant, as COVID-19 vaccines may be less effective in patients infected with *Mycoplasma*.

#### *Do Mycoplasma and COVID-19 Comprise a Binary Biological Weapon?*

The pathogenic *Mycoplasma fermentans incognitus* strain, or Lo's *Mycoplasma*, was patented by Shyh-Ching Lo in 1993 (Patent Number 5,242,820) and several scientists and clinicians have linked this pathogen to over 45% of GWI cases [218]. *This connection was never ruled out, even though Shyh-Ching Lo published in 2000 that there was no serological connection between Mycoplasma fermentans and GWI. However, serological detection of Mycoplasma fermentans is fraught with difficulties that even Lo admits, and his study may have been marred by potential conflict of interest* [219]. Indeed, significant fractions of ME/CFS and FM cases have been associated with *Mycoplasma fermentans* infections as well as those of other *Mycoplasma*

*species, indicating that this pathogen may be involved in various chronic illnesses [220–222].* Thus, the association of long COVID with Mycoplasma infections may establish this bacterial pathogen as a common coinfection in most fatiguing illnesses [175,223]. Moreover, Mycoplasma infections are associated with increased susceptibility to SLE, a condition also associated with excessive fatigue, linking this microorganism to other illnesses marked by exhaustion [224–226]. This could be important, because SLE has been associated with microbial translocation from various niches, possibly linking this autoimmune disease to Mycoplasma colonization [160,175]. Furthermore, lipid-associated membrane proteins (LAMPs) of Mycoplasma fermentans and Mycoplasma hominis have been shown to increase cortisol secretion, further connecting this bacterium to biological barrier dysfunction and chronic fatigue [227]. In 1995, the Institute for Genome Research in Rockville, Maryland completed the nucleotide sequencing of the Mycoplasma genitalium genome, opening the way for the manipulation of this pathogen [228]. Indeed, in 2010, a completely synthetic Mycoplasma mycoides JCVI-syn1.0. was created in the laboratory, contributing further to the potential weaponization of this pathogen [229]. In this regard, Mycoplasma/virus combinations appear suitable for the development of binary biological weapons comprised of independent microorganisms that are considered safe to handle separately, but lethal when mixed, as documented by several studies, including the US Airforce Counterproliferation Center Future Warfare Series No. 53 from 2010 Institute for Molecular Medicine (<https://apps.dtic.mil/sti/pdfs/ADA556597.pdf>, accessed on 29 September 2022) [230,231]. HIV-1 and Mycoplasma fermentans could be an example of this combination. Since SARS-CoV-2/Mycoplasma comorbidity predicts poor COVID-19 prognosis, it is possible that these infections could be further developed as binary biological weapons [232]. Indeed, the SARS-CoV-2 virus is highly contagious and should only be studied in biosafety level 3 (BSL3) laboratories; therefore, the larger scientific community cannot easily study this pathogen or its combinations [233]. Indeed, a microbial cofactor in COVID-19 disease cannot be ruled out, especially since most Mycoplasma species lack reliable antibody tests and are often associated with false-positive COVID-19 serology [193–195]. Moreover, given the possibility of Mycoplasma symbiosis and partnership with other pathogens, including Trichomonas vaginalis, influenza, and HIV, it might be tempting to create a SARS-CoV-2/Mycoplasma coinfection partnership similar to that found in HIV-1 [234–236].

Like SARS viruses, *Mycoplasma species* can also disrupt biological barriers, enabling microbial translocation into host tissues, including the brain. This pathology overlaps with “disorders of unknown etiology” that affect multiple organs and exacerbate many preexistent chronic conditions. Since both *Mycoplasmas* and SARS-CoV-2 induce symptoms that are difficult to connect to a specific etiology, they may be ideal candidates for the development of possible binary biological weapons.

## 5. Interventions

In our previous article on ME/CFS, we introduced some novel treatment strategies for barrier dysfunction, including senotherapy, short chain fatty acids (SCFAs), milk fat globule membranes (MFGM),  $\beta$ -glucan, and fecal microbial transplantation (FMT) [118]. Here, after a short discussion of some senotherapeutic strategies, we introduce HMGB1 inhibitors, TLR4 antagonists, and MLR.

### 5.1. Senotherapeutic Strategies

Senescent cells play a key role in organismal aging, while efferocytosis maintains the homeostasis of biological barriers by clearing senescent or damaged cells. Unchecked accumulation of senescent cells can spread the premature aging phenotype to the neighboring healthy cells via SASP paracrine signaling.

Senotherapeutic agents can be divided into senolytics and senomorphics, the former selectively eliminate senescent cells, while the latter delete the senescent markers p16INK4a and p21CIP1, restoring the cells to pre-senescent status [237,238]. Here, we introduce a third senotherapeutic category, efferocytosis enhancers comprised of: syncytia inhibitors and

blockers of negative efferocytosis regulators. The former subcategory includes TMEM16F inhibitors, while the latter contains inhibitors of anti-efferocytic receptors.

### 5.2. *TMEM16F Inhibitors*

Include drugs like Niclosamide, a widely used anthelmintic agent that inhibits PS externalization, averting both viral fusion with host cells and pathological cell–cell fusion [239,240].

### 5.3. *Negative Efferocytosis Regulator Blockers*

Senescent and cancer cells can avert elimination by expressing CD47, a “don’t eat me” marker that inhibits the key efferocytosis driver, *MER tyrosine kinase* (MERTK), thus blocking the clearance of damaged cells [240–243]. The recently designed CD47 inhibitors, including Hu5F9 and TTI-621, facilitate efferocytosis by blocking the expression of “do not eat me” signals, [244]. These compounds are currently in phase I and II clinical trials, respectively, and are anticipated to receive approval for anticancer indications (NCT04996004 and NCT02216409).

### 5.4. *HMGB1 Antagonists*

In the intracellular compartment, HMGB1 acts as a transcription factor that can facilitate the expression of many genes, including those involved in inflammation and immune responses [245]. Hyperacetylation of HMGB1 causes translocation of this protein from the nucleus into the cytosol where it can act as a danger associated molecular pattern (DAMP), or alarmin [246]. From the cytosol, HMGB1 can be released into the extracellular compartment by disintegrating cells or by secretion from lymphocytes, including NK cells. Extracellular HMGB1 has been associated with illnesses that have fatigue as a major symptom, such as rheumatoid arthritis, atherosclerosis, and certain cancers [245]. HMGB1 attaches to several receptors, including TLR4 and the receptor for advanced glycation end-products (RAGE). Binding to these receptors induces premature cellular senescence in many cell types, including ECs and the result is disruption of endothelial barrier [247–251]. In the GI tract, dysfunctional HMGB1 signaling with RAGE and TLR4, promotes IBD, chronic pain, and the illnesses FM, ME/CFS, long COVID, and GWI (in animal models of this disease) [252–258].

HMGB1 antagonists are likely beneficial for fatigue-related disorders via anti-inflammatory and pro-efferocytic properties. These agents include:

Glycyrrhizin (glycyrrhetic acid), a HMGB1 inhibitor and a traditional medicine, extract from the *Glycyrrhiza glabra* plant, possesses anti-inflammatory, antioxidant, and antimicrobial properties, suggesting it could be beneficial for patients with chronic fatigue [259].

Gabexate mesylate is a synthetic protease inhibitor that blocks HMGB1. This inhibitor showed promising results in preclinical studies, especially for the treatment of neuropathic pain and gut barrier dysfunction [260,261].

Anti-HMGB1 monoclonal antibodies, highly specific antibodies that have been studied for the treatment of several CNS diseases, including stroke, traumatic brain injury (TBI), Parkinson’s disease, epilepsy, and AD, suggesting potentially beneficial results for ME/CFS and similar illnesses [262].

DNA and DNA-like oligonucleotide duplexes, nucleic acids that have been studied in rodents for their anti-inflammatory properties, suggesting a potential role in illnesses marked by inflammation and chronic fatigue [245].

Peptide (HBP08) is a novel pharmacological agent that targets chronic inflammation and fatigue, suggesting that it could be developed as a potential therapy for ME/CFS [263].

N-butanol extracts of *Morinda citrifolia*, that were found to lower intestinal inflammation, and pain in animal models, suggesting that such extracts could be developed for the treatment of chronic fatigue [264–266].

### 5.5. TLR4 Antagonists

TLR4 is a sensor for HMGB1 and LPS, molecules implicated in chronic fatigue, pain, and depression [267]. In addition, TLR4 alters efferocytosis and exacerbates *Mycoplasma* infections, suggesting that biological barriers could be enhanced by inhibiting this protein [268,269] (Table 1). Several TLR4 antagonists are in development as potential therapeutics for IBD, including:

Rhodobacter sphaeroides LPS, a non-toxic molecule that competes with the toxic LPS of Gram-negative bacteria, suggesting a potential benefit as an inhibitor of intestinal barrier disruption [270].

Eritoran (E5564), a synthetic anti-LPS molecule that is considered a second generation TLR4 inhibitor; it has a long duration of action and superior inhibitory properties [271].

TAK-242 is a TLR 4 signaling inhibitor that prevents LPS-induced muscle wasting in mice and probably influences fatigue in humans [272].

**Table 1.** Biological barrier enhancers.

Interventions	Mechanism	References
Senolytic agents	Selective elimination of senescent cells	[237]
Senomorphic agents	Delete senescent markers	[238]
TMEM16F inhibitors	Block PS externalization	[239,240]
CD47 inhibitors	Promote efferocytosis	[244]
HMGB1 antagonists	Inhibit RAGE and TLR4 signaling	[245,246]
TLR4 antagonists	Promote efferocytosis	[270]
MLR	Replace oxidized membrane lipids	[273,274]

### 5.6. Membrane Lipid Replacement (MLR)

SARS-CoV-2-induced cellular senescence causes a phenotype typified by upregulated cytosolic iron which predisposes cells to phospholipid peroxidation of their unsaturated cell membrane glycerolphospholipids and causes ferroptosis [273,274]. Ferroptosis is an iron-induced form of programmed cell death caused by the unchecked accumulation of oxidized lipids in the absence of glutathione peroxidase 4 (GPX4) [275]. Oxidized lipids act as foreign molecules that activate host PRR, triggering chronic inflammation, neuropathic pain, depression, and neurodegeneration [276–278]. Rescue from ferroptotic cell death can be achieved by lowering intracellular iron, increasing GPX4, or replacing cell membrane oxidized lipids. As the SARS-CoV-2 virus upregulates intracellular iron by hijacking host lysosomes and ferritinophagy (ferritin autophagy), restoring cellular iron homeostasis would require lysosomal rehabilitation, a currently unavailable modality. This is illustrated by the paucity of effective treatments for lysosomal disorders [279]. In addition, ferroptotic pores enhance lipid peroxidation by Ca<sup>2+</sup> influx, further lowering GPX4 concentrations [280,281]. Therefore, once activated, ferroptotic cell death takes on a life of its own by initiating a vicious circle of body fat “rusting” and cell death [282]. In addition, enhanced lipid peroxidation and ferroptosis have been associated with ME/CFS, FM, GWI, chronic pain, and neuropsychiatric disorders [283–287]. Interestingly, excess glucocorticoids and ANG II, predispose patients to ferroptosis, while ferroptosis-disintegrating cells release HMGB1, linking this type of programmed cell death to biological barrier dysfunction [288–290].

Natural membrane phospholipid supplementation with fructooligosaccharide-protected glycerolphospholipids, containing unsaturated fatty acids, was demonstrated to safely restore the homeostasis of biological barriers, limiting microbial translocation [291]. The aim of MLR is substitution of ferroptosis-prone polyunsaturated ether phospholipids (PUFA-ePLs) and oxidized lipids with healthy unsaturated glycerolphospholipids [292,293] (Table 1).

## 6. Discussion

The concept of microbial translocation as a key mechanism of chronic systemic immune activation, and disease was studied extensively in the HIV infection, a condition associated with chronic fatigue and increased prevalence of ME/CFS [294,295]. COVID-19, like HIV, causes intestinal barrier disruption, impaired efferocytosis, and accumulation of senescent, apoptotic, and necrotic cells that were previously associated with dysfunctional immune responses [296,297]. Indeed, the newly discovered innate lymphoid cells 3 (ILC3) that release interleukin 22 (IL22), a protector of intestinal barrier, have been implicated in both COVID-19 and HIV, linking dysfunctional mucosal immunity to these viral infections [298,299]. As loss of IL22 was associated with premature cellular senescence, this mechanism may account for the dysfunctional efferocytosis and gut barrier dysfunction in long COVID [300]. Moreover, both IL22 and IL10 protect gut mucosal immunity and act on the same receptors, loss of these cytokines may trigger the pathogenesis of long COVID and ME/CFS [301,302]. These findings are in line not only with our earlier hypothesis but also with the results novel studies that have connected dysfunctional efferocytosis with fatiguing illnesses, including FM, ME/CFS, and GWIs [303–305].

## 7. Conclusions

At the cellular level, life is made possible by cell membranes that separate the intracellular from extracellular compartments and intracellular membranes that separate various organelles from the cell cytoplasm [292]. At the tissue and organismal levels, the gut barrier, comprised of a single layer of epithelial cells, separates luminal prokaryotes from host eukaryotic cells. Although during the development and early life, a limited amount of microbial translocation is thought to help “educate” the immune system to distinguish “self” from “non-self” antigens, later in life gut microbes are immunologically tolerated only in the GI tract.

Weakening of biological barriers and microbial translocation into the systemic circulation, can result in the development of various pathologies, including premature cellular senescence, redox dysfunction, autoimmunity, and elevated inflammatory markers that can be manifested clinically in a variety of forms, such as long COVID, ME/CFS, FMS, GWI, IBD, and even some neuropsychiatric disorders [293]. Ferroptotic signatures, found in these illnesses “of unknown etiology”, point to lipid pathologies, a modifiable risk factor, that may be reversed via novel, strategies, including enhanced clearance of senescent cells, MLR, HMGB1 inhibitors, and TLR4 receptor blockers.

This research connects long COVID to other fatiguing illnesses, including FM, ME/CFS, and GWIs, emphasizing the role of microbial translocation outside the GI tract as the driver of these pathologies. In contrast, correcting the barrier function could ameliorate clinical symptoms as demonstrated in GWIs [293].

**Author Contributions:** Conceptualization, A.S., G.L.N., Z.K. and S.H.; methodology, C.V.A. and L.R.; resources, J.J.A.; writing, A.S. and C.O.; writing—review and editing, C.M.Z.-M.d.C. and J.C.M. All authors have read and agreed to the published version of the manuscript.

**Funding:** This research received no external funding.

**Institutional Review Board Statement:** Not Applicable.

**Informed Consent Statement:** Not Applicable.

**Data Availability Statement:** Not Applicable.

**Conflicts of Interest:** The authors declare no conflict of interest.

## References

1. Raveendran, A.V. Long COVID-19: Challenges in the diagnosis and proposed diagnostic criteria. *Diabetes Metab. Syndr.* **2021**, *15*, 145–146. [[CrossRef](#)] [[PubMed](#)]
2. Shafqat, S.; Arana Chicas, E.; Shafqat, A.; Hashmi, S.K. The Achilles’ heel of cancer survivors: Fundamentals of accelerated cellular senescence. *J. Clin. Investig.* **2022**, *132*, e158452. [[CrossRef](#)] [[PubMed](#)]

3. Ekedahl, H.; Isaksson, S.; Ståhl, O.; Bogefors, K.; Romerius, P.; Eberhard, J.; Giwercman, A. Low-grade inflammation in survivors of childhood cancer and testicular cancer and its association with hypogonadism and metabolic risk factors. *BMC Cancer* **2022**, *22*, 157. [[CrossRef](#)] [[PubMed](#)]
4. Reinertsen, K.V.; Loge, J.H.; Brekke, M.; Kiserud, C.E. Chronic fatigue in adult cancer survivors. *Tidsskr. Nor. Laegeforen.* **2017**, *137*. [[CrossRef](#)]
5. Bøhn, S.H.; Thorsen, L.; Kiserud, C.E.; Fosså, S.D.; Lie, H.C.; Loge, J.H.; Wisløff, T.; Haugnes, H.S.; Reinertsen, K.V. Chronic fatigue and associated factors among long-term survivors of cancers in young adulthood. *Acta Oncol.* **2019**, *58*, 753–762. [[CrossRef](#)]
6. Zheng, G.; Victor Fon, G.; Meixner, W.; Creekmore, A.; Zong, Y.; Dame, M.K.; Colacino, J.; Dedhia, P.H.; Hong, S.; Wiley, J.W. Chronic stress and intestinal barrier dysfunction: Glucocorticoid receptor and transcription repressor HES1 regulate tight junction protein Claudin-1 promoter. *Sci. Rep.* **2017**, *7*, 4502. [[CrossRef](#)]
7. Schinkel, A.H. P-Glycoprotein, a gatekeeper in the blood-brain barrier. *Adv. Drug Deliv. Rev.* **1999**, *36*, 179–194. [[CrossRef](#)]
8. Mason, B.L.; Pariante, C.M.; Thomas, S.A. A revised role for P-glycoprotein in the brain distribution of dexamethasone, cortisol, and corticosterone in wild-type and ABCB1A/B-deficient mice. *Endocrinology* **2008**, *149*, 5244–5253. [[CrossRef](#)]
9. Ueda, K.; Okamura, N.; Hirai, M.; Tanigawara, Y.; Saeki, T.; Kioka, N.; Komano, T.; Hori, R. Human P-glycoprotein transports cortisol, aldosterone, and dexamethasone, but not progesterone. *J. Biol. Chem.* **1992**, *267*, 24248–24252. [[CrossRef](#)]
10. Foley, S.E.; Tuohy, C.; Dunford, M.; Grey, M.J.; De Luca, H.; Cawley, C.; Szabady, R.L.; Maldonado-Contreras, A.; Houghton, J.M.; Ward, D.V.; et al. Gut microbiota regulation of P-glycoprotein in the intestinal epithelium in maintenance of homeostasis. *Microbiome* **2021**, *9*, 183. [[CrossRef](#)]
11. Moriguchi, J.; Kato, R.; Nakagawa, M.; Hirotani, Y.; Ijiri, Y.; Tanaka, K. Effects of lipopolysaccharide on intestinal P-glycoprotein expression and activity. *Eur. J. Pharmacol.* **2007**, *565*, 220–224. [[CrossRef](#)] [[PubMed](#)]
12. Yiallouris, A.; Tsiotis, C.; Agapidaki, E.; Zafeiri, M.; Agouridis, A.P.; Ntourakis, D.; Johnson, E.O. Adrenal Aging and Its Implications on Stress Responsiveness in Humans. *Front Endocrinol (Lausanne)*. *Front. Endocrinol.* **2019**, *10*, 54. [[CrossRef](#)] [[PubMed](#)]
13. Lewis-McDougall, F.C.; Ruchaya, P.J.; Domenjo-Vila, E.; Teoh, T.S.; Prata, L.; Cottle, B.J.; Clark, J.E.; Punjabi, P.P.; Awad, W.; Torella, D.; et al. Aged-senescent cells contribute to impaired heart regeneration. *Aging Cell* **2019**, *18*, e12931. [[CrossRef](#)] [[PubMed](#)]
14. Short, S.; Fielder, E.; Miwa, S.; von Zglinicki, T. Senolytics and senostatics as adjuvant tumour therapy. *eBioMedicine* **2019**, *41*, 683–692. [[CrossRef](#)]
15. Saccon, T.D.; Nagpal, R.; Yadav, H.; Cavalcante, M.B.; Nunes, A.D.C.; Schneider, A.; Gesing, A.; Hughes, B.; Yousefzadeh, M.; Tchkonia, T.; et al. Senolytic Combination of Dasatinib and Quercetin Alleviates Intestinal Senescence and Inflammation and Modulates the Gut Microbiome in Aged Mice. *J. Gerontol. A Biol. Sci. Med. Sci.* **2021**, *76*, 1895–1905. [[CrossRef](#)]
16. Jeyapalan, J.C.; Sedivy, J.M. Cellular senescence and organismal aging. *Mech. Ageing Dev.* **2008**, *129*, 467–474. [[CrossRef](#)]
17. LeBrasseur, N.K.; Tchkonia, T.; Kirkland, J.L. Cellular Senescence and the Biology of Aging, Disease, and Frailty. *Frailty Pathophysiol. Phenotype Patient Care* **2015**, *83*, 11–18. [[CrossRef](#)]
18. Lee, J.-H.; Lee, Y.-K.; Lim, J.J.; Byun, H.-O.; Park, I.; Kim, G.-H.; Xu, W.G.; Wang, H.-J.; Yoon, G. Mitochondrial Respiratory Dysfunction Induces Claudin-1 Expression via Reactive Oxygen Species-mediated Heat Shock Factor 1 Activation, Leading to Hepatoma Cell Invasiveness. *J. Biol. Chem.* **2015**, *290*, 21421–21431. [[CrossRef](#)]
19. Martínez-Cué, C.; Rueda, N. Cellular senescence in neurodegenerative diseases. *Front. Cell Neurosci.* **2020**, *14*, 16. [[CrossRef](#)]
20. Nocerino, A.; Nguyen, A.; Agrawal, M.; Mone, A.; Lakhani, K.; Swaminath, A. Fatigue in Inflammatory Bowel Diseases: Etiologies and Management. *Adv. Ther.* **2020**, *37*, 97–112. [[CrossRef](#)]
21. Martin-Rodriguez, O.; Gauthier, T.; Bonnefoy, F.; Couturier, M.; Daoui, A.; Chagué, C.; Valmary-Degano, S.; Gay, C.; Saas, P.; Perruche, S. Pro-Resolving Factors Released by Macrophages After Efferocytosis Promote Mucosal Wound Healing in Inflammatory Bowel Disease. *Front. Immunol.* **2021**, *12*, 754475. [[CrossRef](#)]
22. Verma, V.; Drury, G.L.; Parisien, M.; Özdağ Acarlı, A.N.; Al-Aubodah, T.A.; Nijnik, A.; Wen, X.; Tugarinov, N.; Verner, M.; Klares, R., 3rd; et al. Unbiased immune profiling reveals a natural killer cell-peripheral nerve axis in fibromyalgia. *Pain* **2022**, *163*, e821–e836. [[CrossRef](#)] [[PubMed](#)]
23. Bi, J. NK cell dysfunction in patients with COVID-19. *Cell Mol. Immunol.* **2022**, *19*, 127–129. [[CrossRef](#)] [[PubMed](#)]
24. Galán, M.; Vigón, L.; Fuertes, D.; Murciano-Antón, M.A.; Casado-Fernández, G.; Domínguez-Mateos, S.; Mateos, E.; Ramos-Martín, F.; Planelles, V.; Torres, M.; et al. Persistent Overactive Cytotoxic Immune Response in a Spanish Cohort of Individuals With Long-COVID: Identification of Diagnostic Biomarkers. *Front. Immunol.* **2022**, *13*, 848886. [[CrossRef](#)]
25. Whistler, T.; Fletcher, M.A.; Lonergan, W.; Zeng, X.R.; Lin, J.M.; Laperrriere, A.; Vernon, S.D.; Klimas, N.G. Impaired immune function in Gulf War Illness. *BMC Med. Genom.* **2009**, *2*, 12. [[CrossRef](#)]
26. Rivas, J.L.; Palencia, T.; Fernández, G.; García, M. Association of T and NK Cell Phenotype With the Diagnosis of Myalgic Encephalomyelitis/Chronic Fatigue Syndrome (ME/CFS). *Front. Immunol.* **2018**, *9*, 1028. [[CrossRef](#)] [[PubMed](#)]
27. Sung, A.P.; Tang, J.J.; Guglielmo, M.J.; Smith-Gagen, J.; Bateman, L.; Navarrete-Galvan, L.; Redelman, D.D.; Hudig, D. Antibody-Dependent Cell-mediated Cytotoxicity (ADCC) in Familial Myalgic Encephalomyelitis/Chronic Fatigue Syndrome (ME/CFS). *Fatigue* **2020**, *8*, 226–244. [[CrossRef](#)] [[PubMed](#)]
28. Lee, E.K.; Sunwoo, J.B. Natural Killer Cells and Thyroid Diseases. *Endocrinol. Metab. Seoul.* **2019**, *34*, 132–137. [[CrossRef](#)] [[PubMed](#)]

29. Bancos, I.; Hazeldine, J.; Chortis, V.; Hampson, P.; Taylor, A.E.; Lord, J.M.; Arlt, W. Primary adrenal insufficiency is associated with impaired natural killer cell function: A potential link to increased mortality. *Eur. J. Endocrinol.* **2017**, *176*, 471–480. [[CrossRef](#)]
30. Lamers, M.M.; Beumer, J.; van der Vaart, J.; Knoops, K.; Puschhof, J.; Breugem, I.T.; Ravelli, R.B.G.; van Schayck, J.P.; Mykytyn, A.Z.; Duimel, H.Q.; et al. SARS-CoV-2 productively infects human gut enterocytes. *Science* **2020**, *369*, 50–54. [[CrossRef](#)]
31. Sanno, N.; Itoh, J.; Teramoto, A.; Itoh, Y.; Hori, S.; Osamura, R.Y. Immunohistochemical detection of human natural killer cell like immunoreactivity in human pituitary adenomas, using monoclonal antibody NK-1. *J. Neurooncol.* **1997**, *35*, 29–38. [[CrossRef](#)] [[PubMed](#)]
32. Belluardo, N.; Mudó, G.; Cella, S.; Santoni, A.; Forni, G.; Bindoni, M. Hypothalamic control of certain aspects of natural immunity in the mouse. *Immunology* **1987**, *62*, 321–327. [[PubMed](#)]
33. Godoy-Pacheco, A.; García-Chagollán, M.; Ramírez-De-Arellano, A.; Hernández-Silva, C.D.; Villegas-Pineda, J.C.; Ramírez-López, I.G.; Zepeda-Nuño, J.S.; Aguilar-Lemarroy, A.; Pereira-Suárez, A.L. Differential modulation of natural killer cell cytotoxicity by 17 $\beta$ -estradiol and prolactin through the NKG2D/NKG2DL axis in cervical cancer cells. *Oncol. Lett.* **2022**, *24*, 288. [[CrossRef](#)] [[PubMed](#)]
34. Mavoungou, E.; Bouyou-Akotet, M.K.; Kremsner, P.G. Effects of prolactin and cortisol on natural killer (NK) cell surface expression and function of human natural cytotoxicity receptors (NKp46, NKp44 and NKp30). *Clin. Exp. Immunol.* **2005**, *139*, 287–296. [[CrossRef](#)] [[PubMed](#)]
35. Jurewicz, M.; McDermott, D.H.; Sechler, J.M.; Tinckam, K.; Takakura, A.; Carpenter, C.B.; Milford, E.; Abdi, R. Human T and natural killer cells possess a functional renin-angiotensin system: Further mechanisms of angiotensin II-induced inflammation. *J. Am. Soc. Nephrol.* **2007**, *18*, 1093–1102. [[CrossRef](#)] [[PubMed](#)]
36. Ambrose, A.R.; Hazime, K.S.; Worboys, J.D.; Niembro-Vivanco, O.; Davis, D.M. Synaptic secretion from human natural killer cells is diverse and includes supramolecular attack particles. *Proc. Natl. Acad. Sci. USA* **2020**, *117*, 23717–23720. [[CrossRef](#)] [[PubMed](#)]
37. Gdynia, G.; Sauer, S.; Kopitz, J.; Fuchs, D.; Duglova, K.; Ruppert, T.; Miller, M.; Pahl, J.; Cerwenka, A.; Enders, M.; et al. The HMGB1 protein induces a metabolic type of tumour cell death by blocking aerobic respiration. *Nat. Commun.* **2016**, *7*, 10764. [[CrossRef](#)]
38. Cerwenka, A.; Kopitz, J.; Schirmacher, P.; Roth, W.; Gdynia, G. HMGB1: The metabolic weapon in the arsenal of NK cells. *Mol. Cell Oncol.* **2016**, *3*, e1175538. [[CrossRef](#)]
39. Peluso, M.J.; Deitchman, A.N.; Torres, L.; Iyer, N.S.; Munter, S.E.; Nixon, C.C.; Donatelli, J.; Thanh, C.; Takahashi, S.; Hakim, J.; et al. Long-term SARS-CoV-2-specific immune and inflammatory responses in individuals recovering from COVID-19 with and without post-acute symptoms. *Cell Rep.* **2021**, *36*, 109518. [[CrossRef](#)]
40. Petruk, G.; Puthia, M.; Petrlova, J.; Samsudin, F.; Strömdahl, A.C.; Cerps, S.; Uller, L.; Kjellström, S.; Bond, P.J.; Schmidtchen, A.A.; et al. SARS-CoV-2 spike protein binds to bacterial lipopolysaccharide and boosts proinflammatory activity. *J. Mol. Cell Biol.* **2020**, *12*, 916–932. [[CrossRef](#)]
41. Štros, M.; Polanská, E.V.; Hlaváčová, T.; Skládal, P. Progress in Assays of HMGB1 Levels in Human Plasma-The Potential Prognostic Value in COVID-19. *Biomolecules* **2022**, *12*, 544. [[CrossRef](#)] [[PubMed](#)]
42. Tripathi, U.; Nchioua, R.; Prata, L.G.P.L.; Zhu, Y.; Gerdes, E.O.W.; Giorgadze, N.; Pirtskhalava, T.; Parker, E.; Xue, A.; Espindola-Netto, J.M.; et al. SARS-CoV-2 causes senescence in human cells and exacerbates the senescence-associated secretory phenotype through TLR-3. *Aging* **2021**, *13*, 21838–21854. [[CrossRef](#)] [[PubMed](#)]
43. Davalos, A.R.; Kawahara, M.; Malhotra, G.K.; Schaum, N.; Huang, J.; Ved, U.; Beausejour, C.M.; Coppe, J.P.; Rodier, F.; Campisi, J. p53-dependent release of Alarmin HMGB1 is a central mediator of senescent phenotypes. *J. Cell Biol.* **2013**, *201*, 613–629. [[CrossRef](#)] [[PubMed](#)]
44. Sofiadis, K.; Josipovic, N.; Nikolic, M.; Kargapolova, Y.; Übelmesser, N.; Varamogianni-Mamatsi, V.; Zirkel, A.; Papadionysiou, I.; Loughran, G.; Keane, J.; et al. HMGB1 coordinates SASP-related chromatin folding and RNA homeostasis on the path to senescence. *Mol. Syst. Biol.* **2021**, *17*, e9760. [[CrossRef](#)]
45. Banerjee, S.; Friggeri, A.; Liu, G.; Abraham, E. The C-terminal acidic tail is responsible for the inhibitory effects of HMGB1 on efferocytosis. *J. Leukoc. Biol.* **2010**, *88*, 973–979. [[CrossRef](#)]
46. Friggeri, A.; Yang, Y.; Banerjee, S.; Park, Y.J.; Liu, G.; Abraham, E. HMGB1 inhibits macrophage activity in efferocytosis through binding to the alphavbeta3-integrin. *Am. J. Physiol. Cell Physiol.* **2010**, *299*, C1267–C1276. [[CrossRef](#)]
47. Nguyen, T.; Johnston, S.; Chacko, A.; Gibson, D.; Cepon, J.; Smith, P.; Staines, D.; Marshall-Gradisnik, S. Novel characterisation of mast cell phenotypes from peripheral blood mononuclear cells in chronic fatigue syndrome/myalgic encephalomyelitis patients. *Asian Pac. J. Allergy Immunol.* **2017**, *35*, 75–81. [[CrossRef](#)]
48. Oktayoglu, P.; Tahtasiz, M.; Bozkurt, M.; Em, S.; Ucar, D.; Yazmalar, L.; Mete, N.; Nas, K.; Gezer, O. Serum levels of high mobility group box 1 protein and its association with quality of life and psychological and functional status in patients with fibromyalgia. *Int. J. Rheum. Dis.* **2013**, *16*, 403–407. [[CrossRef](#)]
49. Garza-Lombó, C.; Thang, M.; Greve, H.J.; Mumaw, C.L.; Messenger, E.J.; Ahmed, C.; Quinn, E.; Sullivan, K.; Block, M.L. Circulating HMGB1 is elevated in veterans with Gulf War Illness and triggers the persistent pro-inflammatory microglia phenotype in male C57Bl/6J mice. *Transl. Psychiatry* **2021**, *11*, 390. [[CrossRef](#)]
50. Hsiao, I.H.; Lin, Y.W. Electroacupuncture Reduces Fibromyalgia Pain by Attenuating the HMGB1, S100B, and TRPV1 Signalling Pathways in the Mouse Brain. *Evid. Based Complement. Altern. Med.* **2022**, *2022*, 2242074. [[CrossRef](#)]

51. Palone, F.; Vitali, R.; Cucchiara, S.; Pierdomenico, M.; Negroni, A.; Aloisio, M.; Nuti, F.; Felice, C.; Armuzzi, A.; Stronati, L. Role of HMGB1 as a suitable biomarker of subclinical intestinal inflammation and mucosal healing in patients with inflammatory bowel disease. *Inflamm. Bowel. Dis.* **2014**, *20*, 1448–1457. [CrossRef] [PubMed]
52. Huang, L.; Zhang, D.; Han, W.; Guo, C. High-mobility group box-1 inhibition stabilizes intestinal permeability through tight junctions in experimental acute necrotizing pancreatitis. *Inflamm. Res.* **2019**, *68*, 677–689. [CrossRef] [PubMed]
53. Zaiatz Bittencourt, V.; Jones, F.; Tosetto, M.; Doherty, G.A.; Ryan, E.J. Dysregulation of Metabolic Pathways in Circulating Natural Killer Cells Isolated from Inflammatory Bowel Disease Patients. *J. Crohns. Colitis.* **2021**, *15*, 1316–1325. [CrossRef] [PubMed]
54. Brogna, C.; Cristoni, S.; Petrillo, M.; Querci, M.; Piazza, O.; Van den Eede, G. Toxin-like peptides in plasma, urine and faecal samples from COVID-19 patients. *F1000Research* **2021**, *10*, 550. [CrossRef]
55. Groff, A.; Kavanaugh, M.; Ramgobin, D.; McClafferty, B.; Aggarwal, C.S.; Golamari, R.; Jain, R. Gastrointestinal Manifestations of COVID-19: A Review of What We Know. *Ochsner J.* **2021**, *21*, 177–180. [CrossRef]
56. Brogna, C.; Brogna, B.; Bisaccia, D.R.; Lauritano, F.; Marino, G.; Montano, L.; Cristoni, S.; Prisco, M.; Piscopo, M. Could SARS-CoV-2 Have Bacteriophage Behavior or Induce the Activity of Other Bacteriophages? *Vaccines* **2022**, *10*, 708. [CrossRef]
57. Aktaş, E.; Özdemir Özgentürk, N. Revealing In Silico that Bacteria's Outer Membrane Proteins may Help our Bodies Replicate and Carry Severe Acute Respiratory Syndrome Coronavirus 2. *Bioinform. Biol. Insights* **2022**, *16*, 1177932221116320. [CrossRef]
58. Ronaldson, P.T.; Davis, T.P. Regulation of blood-brain barrier integrity by microglia in health and disease: A therapeutic opportunity. *J. Cereb. Blood Flow Metab.* **2020**, *40*, S6–S24. [CrossRef]
59. Neal, M.D.; Leaphart, C.; Levy, R.; Prince, J.; Billiar, T.R.; Watkins, S.; Li, J.; Cetin, S.; Ford, H.; Schreiber, A.; et al. Enterocyte TLR4 mediates phagocytosis and translocation of bacteria across the intestinal barrier. *J. Immunol.* **2006**, *176*, 3070–3079. [CrossRef]
60. Lim, J.J.; Grinstein, S.; Roth, Z. Diversity and Versatility of Phagocytosis: Roles in Innate Immunity, Tissue Remodeling, and Homeostasis. *Front. Cell Infect. Microbiol.* **2017**, *7*, 191. [CrossRef]
61. Belizário, J.E.; Neyra, J.M.; Setúbal Destro Rodrigues, M.F. When and how NK cell-induced programmed cell death benefits immunological protection against intracellular pathogen infection. *Innate Immun.* **2018**, *24*, 452–465. [CrossRef] [PubMed]
62. Vann, J.M.; Proctor, R.A. Phagocytosis of bacteria by endothelial cells. In *Pathogenesis of Wound and Biomaterial-Associated Infections*; Wadström, T., Eliasson, I., Holder, I., Ljungh, A., Eds.; Springer: Berlin/Heidelberg, Germany, 1990. [CrossRef]
63. Seeberg, J.C.; Loibl, M.; Moser, F.; Schwegler, M.; Büttner-Herold, M.; Daniel, C.; Engel, F.B.; Hartmann, A.; Schlötzer-Schrehardt, U.; Goppelt-Strübe, M.; et al. Non-professional phagocytosis: A general feature of normal tissue cells. *Sci. Rep.* **2019**, *9*, 11875. [CrossRef] [PubMed]
64. Spits, H.; Artis, D.; Colonna, M.; Diefenbach, A.; Di Santo, J.P.; Eberl, G.; Koyasu, S.; Locksley, R.M.; McKenzie, A.N.; Mebius, R.E.; et al. Innate lymphoid cells—a proposal for uniform nomenclature. *Nat. Rev. Immunol.* **2013**, *13*, 145–149. [CrossRef] [PubMed]
65. Sedgwick, A.J.; Ghazanfari, N.; Constantinescu, P.; Mantamadiotis, T.; Barrow, A.D. The Role of NK Cells and Innate Lymphoid Cells in Brain Cancer. *Front. Immunol.* **2020**, *11*, 1549. [CrossRef]
66. Ermak, G.; Davies, K.J. Calcium and oxidative stress: From cell signaling to cell death. *Mol. Immunol.* **2002**, *38*, 713–721. [CrossRef]
67. Kale, A.; Sharma, A.; Stolzing, A.; Desprez, P.Y.; Campisi, J. Role of immune cells in the removal of deleterious senescent cells. *Immun. Ageing* **2020**, *17*, 16. [CrossRef]
68. Bricogne, C.; Fine, M.; Pereira, P.M.; Sung, J.; Tijani, M.; Wang, Y.; Henriques, R.; Collins, M.K.; Hilgemann, D.W. TMEM16F activation by Ca<sup>2+</sup> triggers plasma membrane expansion and directs PD-1 trafficking. *Sci. Rep.* **2019**, *9*, 619. [CrossRef]
69. Whitlock, J.M.; Chernomordik, L.V. Flagging fusion: Phosphatidylserine signaling in cell-cell fusion. *J. Biol. Chem.* **2021**, *296*, 100411. [CrossRef]
70. Kemp, K.; Wilkins, A.; Scolding, N. Cell fusion in the brain: Two cells forward, one cell back. *Acta Neuropathol.* **2014**, *128*, 629–638. [CrossRef]
71. Schürch, C.M.; Forster, S.; Brühl, F.; Yang, S.H.; Felley-Bosco, E.; Hewer, E. The “don’t eat me” signal CD47 is a novel diagnostic biomarker and potential therapeutic target for diffuse malignant mesothelioma. *Oncoimmunology* **2017**, *7*, e1373235. [CrossRef]
72. Song, P.; An, J.; Zou, M.H. Immune Clearance of Senescent Cells to Combat Ageing and Chronic Diseases. *Cells* **2020**, *9*, 671. [CrossRef] [PubMed]
73. Barrera, L.; Montes-Servín, E.; Hernandez-Martinez, J.M.; García-Vicente, M.L.Á.; Montes-Servín, E.; Herrera-Martínez, M.; Crispín, J.C.; Borbolla-Escoboza, J.R.; Arrieta, O. CD47 overexpression is associated with decreased neutrophil apoptosis/phagocytosis and poor prognosis in non-small-cell lung cancer patients. *Br. J. Cancer* **2017**, *117*, 385–397. [CrossRef] [PubMed]
74. Sweeney, M.D.; Zhao, Z.; Montagne, A.; Nelson, A.R.; Zlokovic, B.V. Blood-Brain Barrier: From Physiology to Disease and Back. *Physiol. Rev.* **2019**, *99*, 21–78. [CrossRef] [PubMed]
75. Spindler, K.R.; Hsu, T.H. Viral disruption of the blood-brain barrier. *Trends Microbiol.* **2012**, *20*, 282–290. [CrossRef]
76. Strazza, M.; Pirrone, V.; Wigdahl, B.; Nonnemacher, M.R. Breaking down the barrier: The effects of HIV-1 on the blood-brain barrier. *Brain Res.* **2011**, *1399*, 96–115. [CrossRef]
77. Diamond, M.S.; Klein, R.S. West Nile virus: Crossing the blood-brain barrier. *Nat. Med.* **2004**, *10*, 1294–1295. [CrossRef]
78. Choi, J.Y.; Park, J.H.; Jo, C.; Kim, K.C.; Koh, Y.H. SARS-CoV-2 spike S1 subunit protein-mediated increase of beta-secretase 1 (BACE1) impairs human brain vessel cells. *Biochem. Biophys. Res. Commun.* **2022**, *626*, 66–71. [CrossRef]
79. Camell, C.D.; Yousefzadeh, M.J.; Zhu, Y.; Prata, L.G.P.L.; Huggins, M.A.; Pierson, M.; Zhang, L.; O’Kelly, R.D.; Pirtskhalava, T.; Xun, P.; et al. Senolytics reduce coronavirus-related mortality in old mice. *Science* **2021**, *373*, eabe4832. [CrossRef]

80. Adesse, D.; Gladulich, L.; Alvarez-Rosa, L.; Siqueira, M.; Marcos, A.C.; Heider, M.; Motta, C.S.; Torices, S.; Toborek, M.; Stipursky, J. Role of aging in Blood–Brain Barrier dysfunction and susceptibility to SARS-CoV-2 infection: Impacts on neurological symptoms of COVID-19. *Fluids Barriers CNS* **2022**, *19*, 63. [[CrossRef](#)]
81. Teixeira, P.C.; Dorneles, G.P.; Filho, P.C.S.; da Silva, I.M.; Schipper, L.L.; Postiga, I.A.; Neves, C.A.M.; Junior, L.C.R.; Peres, A.; de Souto, J.T.; et al. Increased LPS levels coexist with systemic inflammation and result in monocyte activation in severe COVID-19 patients. *Int. Immunopharmacol.* **2021**, *100*, 108125. [[CrossRef](#)]
82. Gomes-Leal, W. Why microglia kill neurons after neural disorders? The friendly fire hypothesis. *Neural. Regen. Res.* **2019**, *14*, 1499–1502. [[CrossRef](#)] [[PubMed](#)]
83. Hornik, T.C.; Neniskyte, U.; Brown, G.C. Inflammation induces multinucleation of Microglia via PKC inhibition of cytokinesis, generating highly phagocytic multinucleated giant cells. *J. Neurochem.* **2014**, *128*, 650–661. [[CrossRef](#)] [[PubMed](#)]
84. Lu, Z.; Liu, S.; Lopes-Virella, M.F.; Wang, Z. LPS and palmitic acid Co-upregulate microglia activation and neuroinflammatory response. *Compr. Psychoneuroendocrinol.* **2021**, *6*, 100048. [[CrossRef](#)] [[PubMed](#)]
85. Tate, W.; Walker, M.; Sweetman, E.; Helliwell, A.; Peppcorn, K.; Edgar, C.; Blair, A.; Chatterjee, A. Molecular Mechanisms of Neuroinflammation in ME/CFS and Long COVID to Sustain Disease and Promote Relapses. *Front. Neurol.* **2022**, *13*, 877772. [[CrossRef](#)]
86. Sencio, V.; Gallerand, A.; Gomes Machado, M.; Deruyter, L.; Heumel, S.; Soulard, D.; Barthelemy, J.; Cuinat, C.; Vieira, A.T.; Barthelemy, A.; et al. Influenza Virus Infection Impairs the Gut’s Barrier Properties and Favors Secondary Enteric Bacterial Infection through Reduced Production of Short-Chain Fatty Acids. *Infect. Immun.* **2021**, *89*, e0073420. [[CrossRef](#)]
87. Salina, A.C.G.; Dos-Santos, D.; Rodrigues, T.S.; Fortes-Rocha, M.; Freitas-Filho, E.G.; Alzamora-Terrel, D.L.; Castro, I.M.S.; Fraga da Silva, T.F.C.; de Lima, M.H.F.; Nascimento, D.C.; et al. Efferocytosis of SARS-CoV-2-infected dying cells impairs macrophage anti-inflammatory functions and clearance of apoptotic cells. *Elife* **2022**, *11*, e74443. [[CrossRef](#)]
88. Liu, S.; Selvaraj, P.; Lien, C.Z.; Nunez, I.A.; Wu, W.W.; Chou, C.K.; Wang, T.T. The PRRA insert at the S1/S2 site modulates cellular tropism of SARS-CoV-2 and ACE2 usage by the closely related Bat RaTG13. *J. Virol.* **2021**, *95*, e01751-20. [[CrossRef](#)]
89. Lee, P.A.; Tullman-Ercek, D.; Georgiou, G. The bacterial twin-arginine translocation pathway. *Annu. Rev. Microbiol.* **2006**, *60*, 373–395. [[CrossRef](#)]
90. Yan, X.; Hu, S.; Yang, Y.; Xu, D.; Li, H.; Liu, W.; He, X.; Li, G.; Cai, W.; Bu, Z. The Twin-Arginine Translocation System Is Important for Stress Resistance and Virulence of Brucella melitensis. *Infect. Immun.* **2020**, *88*, e00389-20. [[CrossRef](#)]
91. Pereyre, S.; Sirand-Pugnet, P.; Beven, L.; Charron, A.; Renaudin, H.; Barré, A.; Avenaud, P.; Jacob, D.; Couloux, A.; Barbe, V.; et al. Life on arginine for Mycoplasma hominis: Clues from its minimal genome and comparison with other human urogenital mycoplasmas. *PLoS Genet.* **2009**, *5*, e1000677. [[CrossRef](#)]
92. Birge, R.B.; Boeltz, S.; Kumar, S.; Carlson, J.; Wanderley, J.; Calianese, D.; Barcinski, M.; Brekken, R.A.; Huang, X.; Hutchins, J.T.; et al. Phosphatidylserine is a global immunosuppressive signal in efferocytosis, infectious disease, and cancer. *Cell Death Differ.* **2016**, *23*, 962–978. [[CrossRef](#)] [[PubMed](#)]
93. Gal, H.; Krizhanovsky, V. Cell fusion induced senescence. *Aging* **2014**, *6*, 353–354. [[CrossRef](#)] [[PubMed](#)]
94. Urata, R.; Ikeda, K.; Yamazaki, E.; Ueno, D.; Katayama, A.; Shin-Ya, M.; Ohgitani, E.; Mazda, O.; Matoba, S. Senescent endothelial cells are predisposed to SARS-CoV-2 infection and subsequent endothelial dysfunction. *Sci. Rep.* **2022**, *12*, 1–9. [[CrossRef](#)] [[PubMed](#)]
95. Rajeevan, M.S.; Murray, J.; Oakley, L.; Lin, J.S.; Unger, E.R. Association of chronic fatigue syndrome with premature telomere attrition. *J. Transl. Med.* **2018**, *16*, 44. [[CrossRef](#)]
96. Zhang, Z.; Zheng, Y.; Niu, Z.; Zhang, B.; Wang, C.; Yao, X.; Peng, H.; Franca, D.N.; Wang, Y.; Zhu, Y.; et al. SARS-CoV-2 spike protein dictates syncytium-mediated lymphocyte elimination. *Cell Death Differ.* **2021**, *28*, 2765–2777. [[CrossRef](#)] [[PubMed](#)]
97. Qiang, S.; Wei, C. Cell-in-cell: An Emerging Player in COVID-19 and Immune Disorders. *Natl. Sci. Open* **2022**, *1*, 20220001. [[CrossRef](#)]
98. Fluge, Ø.; Rekeland, I.G.; Lien, K.; Thürmer, H.; Borchgrevink, P.C.; Schäfer, C.; Sørland, K.; Aßmus, J.; Ktoridou-Valen, I.; Herder, I.; et al. B-Lymphocyte Depletion in Patients With Myalgic Encephalomyelitis/Chronic Fatigue Syndrome: A Randomized, Double-Blind, Placebo-Controlled Trial. *Ann. Trop. Med.* **2019**, *170*, 585–593. [[CrossRef](#)]
99. Sweetman, E.; Kleffmann, T.; Edgar, C.; de Lange, M.; Vallings, R.; Tate, W. A SWATH-MS analysis of Myalgic Encephalomyelitis/Chronic Fatigue Syndrome peripheral blood mononuclear cell proteomes reveals mitochondrial dysfunction. *J. Transl. Med.* **2020**, *18*, 365. [[CrossRef](#)]
100. Hassett, A.L.; Clauw, D.J.; Williams, D.A. Premature Aging in Fibromyalgia. *Curr. Aging Sci.* **2015**, *8*, 178–185. [[CrossRef](#)]
101. Zundel, C.G.; Krengel, M.H.; Heeren, T.; Yee, M.K.; Grasso, C.M.; Janulewicz Lloyd, P.A.; Coughlin, S.S.; Sullivan, K. Rates of Chronic Medical Conditions in 1991, Gulf War Veterans Compared to the General Population. *Int. J. Environ. Res. Public Health* **2019**, *16*, 949. [[CrossRef](#)]
102. Clark, I.A. Chronic cerebral aspects of long COVID, post-stroke syndromes and similar states share their pathogenesis and perispinal etanercept treatment logic. *Pharmacol. Res. Perspect.* **2022**, *10*, e00926, Erratum in: *Pharmacol Res Perspect.* **2022**, *10*, e00942. [[CrossRef](#)]
103. Subramanian, S.; Geng, H.; Tan, X.D. Cell death of intestinal epithelial cells in intestinal diseases. *Sheng Li Xue Bao* **2020**, *72*, 308–324. [[PubMed](#)]

104. Geng, H.; Bu, H.F.; Liu, F.; Wu, L.; Pfeifer, K.; Chou, P.M.; Wang, X.; Sun, J.; Lu, L.; Pandey, A.; et al. In Inflamed Intestinal Tissues and Epithelial Cells, Interleukin 22 Signaling Increases Expression of H19 Long Noncoding RNA, Which Promotes Mucosal Regeneration. *Gastroenterology* **2018**, *155*, 144–155. [CrossRef] [PubMed]
105. Lechuga, S.; Ivanov, A.I. Disruption of the epithelial barrier during intestinal inflammation: Quest for new molecules and mechanisms. *Biochim. Biophys. Acta Mol. Cell Res.* **2017**, *1864*, 1183–1194. [CrossRef] [PubMed]
106. Shankman, L.S.; Fleury, S.T.; Evans, W.B.; Penberthy, K.K.; Arandjelovic, S.; Blumberg, R.S.; Agaisse, H.; Ravichandran, K.S. Efferocytosis by Paneth cells within the intestine. *Curr. Biol.* **2021**, *31*, 2469–2476.e5. [CrossRef]
107. Satoh-Takayama, N.; Vosshenrich, C.A.; Lesjean-Pottier, S.; Sawa, S.; Lochner, M.; Rattis, F.; Mention, J.J.; Thiam, K.; Cerf-Bensussan, N.; Mandelboim, O.; et al. Microbial flora drives interleukin 22 production in intestinal NKp46+ cells that provide innate mucosal immune defense. *Immunity* **2008**, *29*, 958–970. [CrossRef]
108. Wen, S.; Ling, Y.; Yang, W.; Shen, J.; Li, C.; Deng, W.; Liu, W.; Liu, K. Necroptosis is a key mediator of enterocytes loss in intestinal ischaemia/reperfusion injury. *J. Cell Mol. Med.* **2017**, *21*, 432–443. [CrossRef]
109. Mueller, M.; Cima, I.; Noti, M.; Fuhrer, A.; Jakob, S.; Dubuquoy, L.; Schoonjans, K.; Brunner, T. The nuclear receptor LRH-1 critically regulates extra-adrenal glucocorticoid synthesis in the intestine. *J. Exp. Med.* **2006**, *203*, 2057–2062. [CrossRef]
110. Zheng, G.; Wu, S.P.; Hu, Y.; Smith, D.E.; Wiley, J.W.; Hong, S. Corticosterone mediates stress-related increased intestinal permeability in a region-specific manner. *Neurogastroenterol. Motil.* **2013**, *25*, e127–e139. [CrossRef]
111. Cima, I.; Corazza, N.; Dick, B.; Fuhrer, A.; Herren, S.; Jakob, S.; Ayuni, E.; Mueller, C.; Brunner, T. Intestinal epithelial cells synthesize glucocorticoids and regulate T cell activation. *J. Exp. Med.* **2004**, *200*, 1635–1646. [CrossRef]
112. Zhang, Y.; Wang, Y.; Zhou, D.; Zhang, L.S.; Deng, F.X.; Shu, S.; Wang, L.J.; Wu, Y.; Guo, N.; Zhou, J.; et al. Angiotensin II deteriorates advanced atherosclerosis by promoting MerTK cleavage impairing efferocytosis through the AT1R/ROS/p38 MAPK/ADAM17 pathway. *Am. J. Physiol. Cell Physiol.* **2019**, *317*, C776–C787. [CrossRef] [PubMed]
113. Schulte-Schrepping, J.; Reusch, N.; Paclik, D.; Baßler, K.; Schlickeiser, S.; Zhang, B.; Krämer, B.; Krammer, T.; Brumhard, S.; Bonaguro, L.; et al. Severe COVID-19 is marked by a dysregulated myeloid cell compartment. *Cell* **2020**, *182*, 1419–1440.e23. [CrossRef] [PubMed]
114. Miesbach, W. Pathological Role of Angiotensin II in Severe COVID-19. *TH Open* **2020**, *4*, e138–e144. [CrossRef] [PubMed]
115. Kossmann, S.; Schwenk, M.; Hausding, M.; Karbach, S.H.; Schmidgen, M.I.; Brandt, M.; Knorr, M.; Hu, H.; Kröller-Schön, S.; Schönfelder, T.; et al. Angiotensin II-induced vascular dysfunction depends on interferon- $\gamma$ -driven immune cell recruitment and mutual activation of monocytes and NK-cells. *Arterioscler. Thromb. Vasc. Biol.* **2013**, *33*, 1313–1319. [CrossRef] [PubMed]
116. Biancardi, V.C.; Son, S.J.; Ahmadi, S.; Filosa, J.A.; Stern, J.E. Circulating angiotensin II gains access to the hypothalamus and brain stem during hypertension via breakdown of the blood-brain barrier. *Hypertension* **2014**, *63*, 572–579. [CrossRef] [PubMed]
117. Liu, T.J.; Shi, Y.Y.; Wang, E.B.; Zhu, T.; Zhao, Q. AT1R blocker losartan attenuates intestinal epithelial cell apoptosis in a mouse model of Crohn’s disease. *Mol. Med. Rep.* **2016**, *13*, 1156–1162. [CrossRef]
118. Sfera, A.; Osorio, C.; Zapata Martín Del Campo, C.M.; Pereida, S.; Maurer, S.; Maldonado, J.C.; Kozlakidis, Z. Endothelial Senescence and Chronic Fatigue Syndrome, a COVID-19 Based Hypothesis. *Front. Cell Neurosci.* **2021**, *15*, 673217. [CrossRef]
119. Sfera, A.; Osorio, C.; Jafri, N.; Diaz, E.L.; Campo Maldonado, J.E. Intoxication With Endogenous Angiotensin II: A COVID-19 Hypothesis. *Front. Immunol.* **2020**, *11*, 1472. [CrossRef]
120. D’Agnillo, F.; Walters, K.A.; Xiao, Y.; Sheng, Z.M.; Scherler, K.; Park, J.; Gygli, S.; Rosas, L.A.; Sadtler, K.; Kalish, H.; et al. Lung epithelial and endothelial damage, loss of tissue repair, inhibition of fibrinolysis, and cellular senescence in fatal COVID-19. *Sci. Transl. Med.* **2021**, *13*, eabj7790. [CrossRef]
121. Lee, S.; Yu, Y.; Trimpert, J.; Benthani, F.; Mairhofer, M.; Richter-Pechanska, P.; Wyler, E.; Belenki, D.; Kaltenbrunner, S.; Pammer, M.; et al. Virus-induced senescence is a driver and therapeutic target in COVID-19. *Nature* **2021**, *599*, 283–289. [CrossRef]
122. Evangelou, K.; Veroutis, D.; Paschalaki, K.; Foukas, P.G.; Lagopati, N.; Dimitriou, M.; Papaspypopoulos, A.; Konda, B.; Hazapis, O.; Polyzou, A.; et al. Pulmonary infection by SARS-CoV-2 induces senescence accompanied by an inflammatory phenotype in severe COVID-19: Possible implications for viral mutagenesis. *Eur. Respir. J.* **2022**, *60*, 2102951. [CrossRef] [PubMed]
123. Khan, I.; Schmidt, M.O.; Kallakury, B.; Jain, S.; Mehdikhani, S.; Levi, M.; Mendonca, M.; Welch, W.; Riegel, A.T.; Wilcox, C.S.; et al. Low Dose Chronic Angiotensin II Induces Selective Senescence of Kidney Endothelial Cells. *Front. Cell Dev. Biol.* **2021**, *9*, 782841. [CrossRef] [PubMed]
124. Lekva, T.; Ueland, T.; Halvorsen, B.; Murphy, S.L.; Dyrhol-Riise, A.M.; Tveita, A.; Finbråten, A.K.; Mathiessen, A.; Müller, K.E.; Aaløkken, T.M.; et al. Markers of cellular senescence is associated with persistent pulmonary pathology after COVID-19 infection. *Infect. Dis.* **2022**, *19*, 1–6. [CrossRef] [PubMed]
125. Venzon, M.; Bernard-Raichon, L.; Klein, J.; Axelrad, J.E.; Zhang, C.; Hussey, G.A.; Sullivan, A.P.; Casanovas-Massana, A.; Noval, M.G.; Valero-Jimenez, A.M.; et al. Gut microbiome dysbiosis during COVID-19 is associated with increased risk for bacteremia and microbial translocation. *Biorxiv* **2022**. [CrossRef]
126. Oliva, A.; Miele, M.C.; Di Timoteo, F.; De Angelis, M.; Mauro, V.; Aronica, R.; Al Ismail, D.; Ceccarelli, G.; Pinacchio, C.; d’Ettorre, G.; et al. Persistent Systemic Microbial Translocation and Intestinal Damage During Coronavirus Disease-19. *Front. Immunol.* **2021**, *12*, 708149. [CrossRef] [PubMed]
127. Giron, L.B.; Peluso, M.J.; Ding, J.; Kenny, G.; Zilberman, N.F.; Koshy, J.; Hong, K.Y.; Rasmussen, H.; Miller, G.E.; Bishehsari, F.; et al. Markers of fungal translocation are elevated during post-acute sequelae of SARS-CoV-2 and induce NF- $\kappa$ B signaling. *JCI Insight* **2022**, *7*, e160989. [CrossRef] [PubMed]

128. Kates, A.; Keating, J.; Baubie, K.; Putman-Buehler, N.; Watson, L.; Godfrey, J.; Deblois, C.L.; Suen, G.; Cook, D.B.; Rabago, D.; et al. Examining the association between the gastrointestinal microbiota and Gulf War illness: A prospective cohort study. *PLoS ONE* **2022**, *17*, e0268479. [[CrossRef](#)]
129. Minerbi, A.; Fitzcharles, M.A. Gut microbiome: Pertinence in fibromyalgia. *Clin. Exp. Rheumatol.* **2020**, *38* (Suppl. S123), 99–104.
130. Zhang, B.; Verne, M.L.; Fields, J.Z.; Verne, G.N.; Zhou, Q. Intestinal Hyperpermeability in Gulf War Veterans With Chronic Gastrointestinal Symptoms. *J. Clin. Gastroenterol.* **2019**, *53*, e298–e302. [[CrossRef](#)]
131. Fine, R.L.; Manfredo Vieira, S.; Gilmore, M.S.; Kriegel, M.A. Mechanisms and consequences of gut commensal translocation in chronic diseases. *Gut Microbes* **2020**, *11*, 217–230. [[CrossRef](#)]
132. Kouzu, K.; Tsujimoto, H.; Kishi, Y.; Ueno, H.; Shinomiya, N. Bacterial Translocation in Gastrointestinal Cancers and Cancer Treatment. *Biomedicines* **2022**, *10*, 380. [[CrossRef](#)]
133. Zhu, F.; Ju, Y.; Wang, W.; Wang, Q.; Guo, R.; Ma, Q.; Sun, Q.; Fan, Y.; Xie, Y.; Yang, Z.; et al. Metagenome-wide association of gut microbiome features for schizophrenia. *Nat. Commun.* **2020**, *11*, 1–10. [[CrossRef](#)] [[PubMed](#)]
134. Stamova, B.; Sharp, F.R. Lipopolysaccharide Associates with Amyloid Plaques, Neurons and Oligodendrocytes in Alzheimer’s Disease Brain: A Review. *Front. Aging Neurosci.* **2018**, *10*, 42. [[CrossRef](#)]
135. Clarke, M.B.; Hughes, D.T.; Zhu, C.; Boedeker, E.C.; Sperandio, V. The QseC sensor kinase: A bacterial adrenergic receptor. *Proc. Natl. Acad. Sci. USA* **2006**, *103*, 10420–10425. [[CrossRef](#)] [[PubMed](#)]
136. Patel, P.A.; Hernandez, A.F. Targeting anti-beta-1-adrenergic receptor antibodies for dilated cardiomyopathy. *Eur. J. Heart Fail.* **2013**, *15*, 724–729. [[CrossRef](#)] [[PubMed](#)]
137. Loebel, M.; Grabowski, P.; Heidecke, H.; Bauer, S.; Hanitsch, L.G.; Wittke, K.; Meisel, C.; Reinke, P.; Volk, H.D.; Fluge, Ø.; et al. Antibodies to β adrenergic and muscarinic cholinergic receptors in patients with Chronic Fatigue Syndrome. *Brain Behav. Immun.* **2016**, *52*, 32–39. [[CrossRef](#)]
138. Wessler, I.K.; Kirkpatrick, C.J. Non-neuronal acetylcholine involved in reproduction in mammals and honeybees. *J. Neurochem.* **2017**, *142* (Suppl. S2), 144–150. [[CrossRef](#)]
139. Bynke, A.; Julin, P.; Gottfries, C.G.; Heidecke, H.; Scheibenbogen, C.; Bergquist, J. Autoantibodies to beta-adrenergic and muscarinic cholinergic receptors in Myalgic Encephalomyelitis (ME) patients—A validation study in plasma and cerebrospinal fluid from two Swedish cohorts. *Brain Behav. Immun. Health* **2020**, *7*, 100107. [[CrossRef](#)]
140. Iwasa, K.; Yoshikawa, H.; Hamaguchi, T.; Sakai, K.; Shinohara-Noguchi, M.; Samuraki, M.; Takahashi, K.; Yanase, D.; Ono, K.; Ishida, C.; et al. Time-series analysis: Variation of anti-acetylcholine receptor antibody titer in myasthenia gravis is related to incidence of Mycoplasma pneumoniae and influenza virus infections. *Neurol. Res.* **2018**, *40*, 102–109. [[CrossRef](#)]
141. Elsaïs, A.; Wyller, V.B.; Loge, J.H.; Kerty, E. Fatigue in myasthenia gravis: Is it more than muscular weakness? *BMC Neurol.* **2013**, *13*, 132. [[CrossRef](#)]
142. Alekseeva, T.M.; Gavrilov, Y.V.; Kreis, O.A.; Valko, P.O.; Weber, K.P.; Valko, Y. Fatigue in patients with myasthenia gravis. *J. Neurol.* **2018**, *265*, 2312–2321. [[CrossRef](#)] [[PubMed](#)]
143. Bjordan, T.; Mehl, T.L.K.; Schweden, U.; Menge, S. Assessment of physical fatigability and fatigue perception in myasthenia gravis. *Muscle Nerve* **2017**, *55*, 657–663. [[CrossRef](#)] [[PubMed](#)]
144. Ruiter, A.M.; Verschuur, J.J.G.M.; Tannemaat, M.R. Fatigue in patients with myasthenia gravis. A systematic review of the literature. *Neuromuscul. Disord.* **2020**, *30*, 631–639. [[CrossRef](#)] [[PubMed](#)]
145. Gable, M.S.; Gavali, S.; Radner, A.; Tilley, D.H.; Lee, B.; Dyner, L.; Collins, A.; Dengel, A.; Dalmau, J.; Glaser, C.A. Anti-NMDA receptor encephalitis: Report of ten cases and comparison with viral encephalitis. *Eur. J. Clin. Microbiol. Infect. Dis.* **2009**, *28*, 1421–1429. [[CrossRef](#)] [[PubMed](#)]
146. Tong, J.; Huang, J.; Luo, X.; Chen, S.; Cui, Y.; An, H.; Xiu, M.; Tan, S.; Wang, Z.; Yuan, Y.; et al. Elevated serum anti-NMDA receptor antibody levels in first-episode patients with schizophrenia. *Brain Behav. Immun.* **2019**, *81*, 213–219. [[CrossRef](#)]
147. Wang, J.; Yang, G.; Wang, X.; Wen, Z.; Shuai, L.; Luo, J.; Wang, C.; Sun, Z.; Liu, R.; Ge, J.; et al. SARS-CoV-2 uses metabotropic glutamate receptor subtype 2 as an internalization factor to infect cells. *Cell Discov.* **2021**, *7*, 119. [[CrossRef](#)]
148. Wang, J.; Wang, Z.; Liu, R.; Shuai, L.; Wang, X.; Luo, J.; Wang, C.; Chen, W.; Wang, X.; Ge, J.; et al. Metabotropic glutamate receptor subtype 2 is a cellular receptor for rabies virus. *PLoS Pathog.* **2018**, *14*, e1007189. [[CrossRef](#)]
149. Ruiz-García, R.; Martínez-Hernández, E.; Joubert, B.; Petit-Pedrol, M.; Pajarón-Boix, E.; Fernández, V.; Salais, L.; Del Pozo, M.; Armangué, T.; Sabater, L.; et al. Paraneoplastic cerebellar ataxia and antibodies to metabotropic glutamate receptor 2. *Neurol. Neuroimmunol. Neuroinflamm.* **2019**, *7*, e658. [[CrossRef](#)]
150. Mazzitelli, M.; Palazzo, E.; Maione, S.; Neugebauer, V. Group II Metabotropic Glutamate Receptors: Role in Pain Mechanisms and Pain Modulation. *Front. Mol. Neurosci.* **2018**, *11*, 383. [[CrossRef](#)]
151. Dalmau, J.; Geis, C.; Graus, F. Autoantibodies to Synaptic Receptors and Neuronal Cell Surface Proteins in Autoimmune Diseases of the Central Nervous System. *Physiol. Rev.* **2017**, *97*, 839–887. [[CrossRef](#)]
152. Thye, A.Y.; Law, J.W.; Tan, L.T.; Thurairajasingam, S.; Chan, K.G.; Letchumanan, V.; Lee, L.H. Exploring the Gut Microbiome in Myasthenia Gravis. *Nutrients* **2022**, *14*, 1647. [[CrossRef](#)] [[PubMed](#)]
153. Qiu, D.; Xia, Z.; Jiao, X.; Deng, J.; Zhang, L.; Li, J. Altered Gut Microbiota in Myasthenia Gravis. *Front. Microbiol.* **2018**, *9*, 2627. [[CrossRef](#)] [[PubMed](#)]
154. Scarpellini, E.; Ianiro, G.; Attili, F.; Bassanelli, C.; De Santis, A.; Gasbarrini, A. The human gut microbiota and virome: Potential therapeutic implications. *Dig. Liver Dis.* **2015**, *47*, 1007–1012. [[CrossRef](#)] [[PubMed](#)]

155. Robinson, C.M.; Pfeiffer, J.K. Viruses and the Microbiota. *Annu. Rev. Virol.* **2014**, *1*, 55–69. [CrossRef] [PubMed]
156. Guerin, E.; Hill, C. Shining Light on Human Gut Bacteriophages. *Front. Cell Infect. Microbiol.* **2020**, *10*, 481. [CrossRef] [PubMed]
157. Petrillo, M.; Brogna, C.; Cristoni, S.; Querci, M.; Piazza, O.; Van den Eede, G. Increase of SARS-CoV-2 RNA load in faecal samples prompts for rethinking of SARS-CoV-2 biology and COVID-19 epidemiology. *F1000Research* **2021**, *10*, 370. [CrossRef]
158. Vishnyakov, I.E. Cell-in-Cell Phenomena in Wall-Less Bacteria: Is It Possible? *Int. J. Mol. Sci.* **2022**, *23*, 4345. [CrossRef]
159. Gold, J.E.; Okyay, R.A.; Licht, W.E.; Hurley, D.J. Investigation of Long COVID Prevalence and Its Relationship to Epstein-Barr Virus Reactivation. *Pathogens* **2021**, *10*, 763. [CrossRef]
160. Ma, L.; Morel, L. Loss of Gut Barrier Integrity In Lupus. *Front. Immunol.* **2022**, *13*, 919792. [CrossRef]
161. Ogunrinde, E.; Zhou, Z.; Luo, Z.; Alekseyenko, A.; Li, Q.Z.; Macedo, D.; Kamen, D.L.; Oates, J.C.; Gilkeson, G.S.; Jiang, W. A Link Between Plasma Microbial Translocation, Microbiome, and Autoantibody Development in First-Degree Relatives of Systemic Lupus Erythematosus Patients. *Arthritis Rheumatol.* **2019**, *71*, 1858–1868. [CrossRef]
162. Carneiro-Filho, B.A.; Lima, I.P.; Araujo, D.H.; Cavalcante, M.C.; Carvalho, G.H.; Brito, G.A.; Lima, V.; Monteiro, S.M.N.; Santos, F.N.; Ribeiro, R.A.; et al. Intestinal barrier function and secretion in methotrexate-induced rat intestinal mucositis. *Dig. Dis. Sci.* **2004**, *49*, 65–72. [CrossRef] [PubMed]
163. Song, D.; Shi, B.; Xue, H.; Li, Y.; Yang, X.; Yu, B.; Xu, Z.; Liu, F.; Li, J. Confirmation and prevention of intestinal barrier dysfunction and bacterial translocation caused by methotrexate. *Dig. Dis. Sci.* **2006**, *51*, 1549–1556. [CrossRef] [PubMed]
164. De Arumahandi Silva, A.N.; Frommert, L.M.; Albach, F.N.; Klotsche, J.; Scholz, V.; Jeworowski, L.M.; Schwarz, T.; Ten Hagen, A.; Zernicke, J.; Corman, V.M.; et al. Pausing methotrexate improves immunogenicity of COVID-19 vaccination in elderly patients with rheumatic diseases. *Ann. Rheum. Dis.* **2022**, *81*, 881–888. [CrossRef] [PubMed]
165. Razin, S.; Yogeve, D.; Naot, Y. Molecular biology and pathogenicity of mycoplasmas. *Microbiol. Mol. Biol. Rev.* **1998**, *62*, 1094–1156. [CrossRef]
166. Fadiel, A.; Eichenbaum, K.D.; Semary, N.E.; Epperson, B. Mycoplasma genomics: Tailoring the genome for minimal life requirements through reductive evolution. *Front. Biosci.* **2007**, *12*, 2020–2028. [CrossRef]
167. Glass, J.I.; Assad-Garcia, N.; Alperovich, N.; Yooseph, S.; Lewis, M.R.; Maruf, M.; Hutchison, C.A.; Smith, H.O.; Venter, J.C. Essential genes of a minimal bacterium. *Proc. Natl. Acad. Sci. USA* **2006**, *103*, 425–430. [CrossRef]
168. Rawadi, G.; Roman-Roman, S. Mycoplasma membrane lipoproteins induced proinflammatory cytokines by a mechanism distinct from that of lipopolysaccharide. *Infect. Immun.* **1996**, *64*, 637–643. [CrossRef]
169. Kotecha, A.; Wang, Q.; Dong, X.; Ilca, S.L.; Ondivieila, M.; Zihe, R.; Seago, J.; Charleston, B.; Fry, E.E.; Abrescia, N.G.; et al. Rules of engagement between  $\alpha\beta\delta$  integrin and foot-and-mouth disease virus. *Nat. Commun.* **2017**, *8*, 1–8. [CrossRef]
170. Fan, W.; Qian, P.; Wang, D.; Zhi, X.; Wei, Y.; Chen, H.; Li, X. Integrin  $\alpha\beta\delta$  promotes infection by Japanese encephalitis virus. *Res. Vet. Sci.* **2017**, *111*, 67–74. [CrossRef]
171. He, Q.Q.; Ren, S.; Xia, Z.C.; Cheng, Z.K.; Peng, N.F.; Zhu, Y. Fibronectin facilitates enterovirus 71 infection by mediating viral entry. *J. Virol.* **2018**, *92*, e02251-17. [CrossRef]
172. Rechnitzer, H.; Rottem, S.; Herrmann, R. Reconstitution of an active arginine deiminase pathway in *Mycoplasma pneumoniae* M129. *Infect. Immun.* **2013**, *81*, 3742–3749. [CrossRef] [PubMed]
173. Huang, Z.; Hu, H. Arginine Deiminase Induces Immunogenic Cell Death and Is Enhanced by N-acetylcysteine in Murine MC38 Colorectal Cancer Cells and MDA-MB-231 Human Breast Cancer Cells In Vitro. *Molecules* **2021**, *26*, 511. [CrossRef] [PubMed]
174. Nicolson, G.L.; de Mattos, G.F. COVID-19 Coronavirus: Is Infection along with *Mycoplasma* or Other Bacteria Linked to Progression to a Lethal Outcome? *Int. J. Clin. Med.* **2020**, *11*, 5. [CrossRef]
175. Gayam, V.; Konala, V.M.; Naramala, S.; Garlapati, P.R.; Merghani, M.A.; Regmi, N.; Balla, M.; Adapa, S. Presenting characteristics, comorbidities, and outcomes of patients coinfected with COVID-19 and *Mycoplasma pneumoniae* in the USA. *J. Med. Virol.* **2020**, *92*, 2181–2187. [CrossRef] [PubMed]
176. Choubey, A.; Sagar, D.; Cawley, P.; Miller, K. Retrospective review analysis of COVID-19 patients co-infected with *Mycoplasma pneumoniae*. *Lung India* **2021**, *38* Suppl. S1, S22–S26. [CrossRef] [PubMed]
177. Rangroo, R.; Young, M.; Davis, A.; Pack, S.; Thakore, S.; Schepcoff, A.; Oyesanmi, O. The Severity of the Co-infection of *Mycoplasma pneumoniae* in COVID-19 Patients. *Cureus* **2022**, *14*, e24563. [CrossRef]
178. Cai, F.; Shou, X.; Ye, Q. Epidemiological Study on *Mycoplasma pneumoniae* and *Chlamydia pneumoniae* Infection of Hospitalized Children in a Single Center During the COVID-19 Pandemic. *Front. Cell. Infect. Microbiol.* **2022**, *12*, 843463. [CrossRef]
179. Lai, C.C.; Wang, C.Y.; Hsueh, P.R. Co-infections among patients with COVID-19: The need for combination therapy with non-anti-SARS-CoV-2 agents? *J. Microbiol. Immunol. Infect.* **2020**, *53*, 505–512. [CrossRef]
180. Nicolson, G.L.; Haier, J.; Nasralla, M.; Nicolson, N.L.; Ngwenya, R.; De Meirleir, K. Mycoplasmal infections in Chronic Fatigue Syndrome, Fibromyalgia Syndrome and Gulf War Illness. *J. Chron. Fatigue Syndr.* **2000**, *6*, 23–39. [CrossRef]
181. Chen, W.; Li, D.; Paulus, B.; Wilson, I.; Chadwick, V.S. High prevalence of *Mycoplasma pneumoniae* in intestinal mucosal biopsies from patients with inflammatory bowel disease and controls. *Dig. Dis. Sci.* **2001**, *46*, 2529–2535. [CrossRef]
182. Cortes, G.M.; Marcialis, M.A.; Bardanzellu, F.; Corrias, A.; Fanos, V.; Mussap, M. Inflammatory Bowel Disease and COVID-19: How Microbiomics and Metabolomics Depict Two Sides of the Same Coin. *Front. Microbiol.* **2022**, *13*, 856165. [CrossRef] [PubMed]
183. Tsiodras, S.; Kelesidis, I.; Kelesidis, T.; Stamboulis, E.; Giannarellou, H. Central nervous system manifestations of *Mycoplasma pneumoniae* infections. *J. Infect.* **2005**, *51*, 343–354. [CrossRef] [PubMed]

184. Choo, Q.W.W.; Koean, R.A.G.; Chang, S.C.; Chng, W.J.; Chan, M.C.; Wang, W.; Er, J.Z.; Ding, J.L. Macrophages protect mycoplasma-infected chronic myeloid leukemia cells from natural killer cell killing. *Immunol. Cell Biol.* **2020**, *98*, 138–151, Epub 2020. [CrossRef] [PubMed]
185. Blanchard, A.; Montagnier, L. AIDS-associated mycoplasmas. *Annu. Rev. Microbiol.* **1994**, *48*, 687–712. [CrossRef] [PubMed]
186. Mina, M.J.; Burke, R.M.; Klugman, K.P. Estimating the prevalence of coinfection with influenza virus and the atypical bacteria *Bordetella pertussis*, *Chlamydophila pneumoniae*, and *Mycoplasma pneumoniae*. *Eur. J. Clin. Microbiol. Infect. Dis.* **2014**, *33*, 1585–1589. [CrossRef]
187. Huang, S.; Li, J.Y.; Wu, J.; Meng, L.; Shou, C.C. Mycoplasma infections and different human carcinomas. *World J. Gastroenterol.* **2001**, *7*, 266–269. [CrossRef]
188. Monte Serrano, J.; García-Gil, M.F.; Cruañes Monferrer, J.; Aldea Manrique, B.; Prieto-Torres, L.; García García, M.; Matovelle Ochoa, C.; Ara-Martín, M. COVID-19 and Mycoplasma pneumoniae: SARS-CoV-2 false positive or coinfection? *Int. J. Dermatol.* **2020**, *59*, 1282–1283. [CrossRef]
189. Chaudhry, R.; Sreenath, K.; Vinayaraj, E.V.; Sahoo, B.; Vishnu Narayanan, M.R.; Kiran, K.V.P.S.; Batra, P.; Rathor, N.; Singh, S.; Mohan, A.; et al. Mycoplasma pneumoniae co-infection with SARS-CoV-2: A case report. *Access Microbiol.* **2021**, *3*, 000212. [CrossRef]
190. Oliva, A.; Siccardi, G.; Migliarini, A.; Cancelli, F.; Carnevalini, M.; D'Andria, M.; Attilia, I.; Danese, V.C.; Cecchetti, V.; Romiti, R.; et al. Co-infection of SARS-CoV-2 with Chlamydia or Mycoplasma pneumoniae: A case series and review of the literature. *Infection* **2020**, *48*, 871–877. [CrossRef]
191. Waites, K.B.; Talkington, D.F. Mycoplasma pneumoniae and its role as a human pathogen. *Clin. Microbiol. Rev.* **2004**, *17*, 697–728, table of contents. [CrossRef]
192. Loens, K.; Ursi, D.; Goossens, H.; Ieven, M. Molecular diagnosis of Mycoplasma pneumoniae respiratory tract infections. *J. Clin. Microbiol.* **2003**, *41*, 4915–4923. [CrossRef] [PubMed]
193. Shen, H.; Zhu, B.; Wang, S.; Mo, H.; Wang, J.; Li, J.; Zhang, C.; Zeng, H.; Guan, L.; Shi, W.; et al. Association of targeted multiplex PCR with resequencing microarray for the detection of multiple respiratory pathogens. *Front. Microbiol.* **2015**, *6*, 532. [CrossRef] [PubMed]
194. Halbedel, S.; Stölke, J. Tools for the genetic analysis of Mycoplasma. *Int. J. Med. Microbiol.* **2007**, *297*, 37–44. [CrossRef] [PubMed]
195. Huber, T.; Steininger, P.; Irrgang, P.; Korn, K.; Tenbusch, M.; Diesch, K.; Achenbach, S.; Kremer, A.E.; Werblow, M.; Vetter, M.; et al. Diagnostic performance of four SARS-CoV-2 antibody assays in patients with COVID-19 or with bacterial and non-SARS-CoV-2 viral respiratory infections. *Eur. J. Clin. Microbiol. Infect. Dis.* **2021**, *40*, 1983–1997. [CrossRef]
196. Li, N.; Ma, X.; Zhou, J.; Deng, J.; Gu, C.; Fei, C.; Cao, L.; Zhang, Q.; Tao, F. Clinical application of metagenomic next-generation sequencing technology in the diagnosis and treatment of pulmonary infection pathogens: A prospective single-center study of 138 patients. *J. Clin. Lab. Anal.* **2022**, *36*, e24498. [CrossRef]
197. Martire, B.; Foti, C.; Cassano, N.; Buquicchio, R.; Del Vecchio, G.C.; De Mattia, D. Persistent B-cell lymphopenia, multiorgan disease, and erythema multiforme caused by Mycoplasma pneumoniae infection. *Pediatr. Dermatol.* **2005**, *22*, 558–560. [CrossRef]
198. Zhao, Q.; Meng, M.; Kumar, R.; Wu, Y.; Huang, J.; Deng, Y.; Weng, Z.; Yang, L. Lymphopenia is associated with severe coronavirus disease 2019, (COVID-19) infections: A systemic review and meta-analysis. *Int. J. Infect. Dis.* **2020**, *96*, 131–135. [CrossRef]
199. Mirijello, A.; La Marca, A.; D'Errico, M.M.; Curci, S.; Vendemiale, G.; Grandone, E.; De Cosmo, S. Venous thromboembolism during mycoplasma pneumoniae infection: Case report and review of the literature. *Eur. Rev. Med. Pharmacol. Sci.* **2020**, *24*, 10061–10068. [CrossRef]
200. Porfidia, A.; Valeriani, E.; Pola, R.; Porreca, E.; Rutjes, A.W.S.; Di Nisio, M. Venous thromboembolism in patients with COVID-19: Systematic review and meta-analysis. *Thromb. Res.* **2020**, *196*, 67–74. [CrossRef]
201. Ali, A.S.; ASattar, M.A.; Karim, S.; Kutbi, D.; Aljohani, H.; Bakhshwin, D.; Alsieni, M.; Alkreathy, H.M. Pharmacological basis for the potential role of Azithromycin and Doxycycline in management of COVID-19. *Arab. J. Chem.* **2021**, *14*, 102983. [CrossRef]
202. Kouegnigan Rerambiah, L.; Ndong, J.C.; Medzegue, S.; Elisee-Ndam, M.; Djoba Siawaya, J.F. Genital Mycoplasma infections and their resistance phenotypes in an African setting. *Eur. J. Clin. Microbiol. Infect. Dis.* **2015**, *34*, 1087–1090. [CrossRef] [PubMed]
203. Ruoslahti, E. RGD and other recognition sequences for integrins. *Annu. Rev. Cell Dev. Biol.* **1996**, *12*, 697–715. [CrossRef] [PubMed]
204. May, M.; Papazisi, L.; Gorton, T.S.; Geary, S.J. Identification of fibronectin-binding proteins in *Mycoplasma gallisepticum* strain R. *Infect. Immun.* **2006**, *74*, 1777–1785. [CrossRef] [PubMed]
205. Widjaja, M.; Berry, I.J.; Jarocki, V.M.; Padula, M.P.; Dumke, R.; Djordjevic, S.P. Cell surface processing of the P1 adhesin of Mycoplasma pneumoniae identifies novel domains that bind host molecules. *Sci. Rep.* **2020**, *10*, 1–16. [CrossRef] [PubMed]
206. Zimmermann, L.; Peterhans, E.; Frey, J. RGD motif of lipoprotein T, involved in adhesion of *Mycoplasma conjunctivae* to lamb synovial tissue cells. *J. Bacteriol.* **2010**, *192*, 3773–3779. [CrossRef] [PubMed]
207. Liu, J.; Lu, F.; Chen, Y.; Plow, E.; Qin, J. Integrin mediates cell entry of the SARS-CoV-2 virus independent of cellular receptor ACE2. *J. Biol. Chem.* **2022**, *298*, 101710. [CrossRef]
208. Nader, D.; Fletcher, N.; Curley, G.F.; Kerrigan, S.W. SARS-CoV-2 uses major endothelial integrin  $\alpha v\beta 3$  to cause vascular dysregulation in-vitro during COVID-19. *PLoS ONE* **2021**, *16*, e0253347. [CrossRef]
209. Lemańska-Perek, A.; Krzyżanowska-Gołąb, D.; Dragan, B.; Tyszko, M.; Adamik, B. Fibronectin as a Marker of Disease Severity in Critically Ill COVID-19 Patients. *Cells* **2022**, *11*, 1566. [CrossRef]

210. Balasubramanian, S.; Kannan, T.R.; Baseman, J.B. The surface-exposed carboxyl region of *Mycoplasma pneumoniae* elongation factor Tu interacts with fibronectin. *Infect. Immun.* **2008**, *76*, 3116–3123. [CrossRef]
211. Tarshis, M.; Morag, B.; Mayer, M. *Mycoplasma* cells stimulate in vitro activation of plasminogen by purified tissue-type plasminogen activator. *FEMS Microbiol. Lett.* **1993**, *106*, 201–204. [CrossRef]
212. Yavlovich, A.; Higazi, A.A.; Rottem, S. Plasminogen binding and activation by *Mycoplasma fermentans*. *Infect. Immun.* **2001**, *69*, 1977–1982. [CrossRef] [PubMed]
213. Ji, H.L.; Zhao, R.; Matalon, S.; Matthay, M.A. Elevated Plasmin(ogen) as a Common Risk Factor for COVID-19 Susceptibility. *Physiol. Rev.* **2020**, *100*, 1065–1075. [CrossRef] [PubMed]
214. Berlanga-Acosta, J.A.; Guillén-Nieto, G.E.; Rodríguez-Rodríguez, N.; Mendoza-Mari, Y.; Bringas-Vega, M.L.; Berlanga-Saez, J.O.; García Del Barco Herrera, D.; Martínez-Jimenez, I.; Hernandez-Gutierrez, S.; Valdés-Sosa, P.A. Cellular Senescence as the Pathogenic Hub of Diabetes-Related Wound Chronicity. *Front. Endocrinol.* **2020**, *11*, 573032. [CrossRef]
215. Dowsett, J.; Ferklingstad, E.; Rasmussen, L.J.H.; Thørner, L.W.; Magnússon, M.K.; Sugden, K.; Thorleifsson, G.; Frigge, M.; Burgdorf, K.S.; Ostrowski, S.R.; et al. Eleven genomic loci affect plasma levels of chronic inflammation marker soluble urokinase-type plasminogen activator receptor. *Commun. Biol.* **2021**, *4*, 655. [CrossRef] [PubMed]
216. Arfi, Y.; Lartigue, C.; Sirand-Pugnet, P.; Blanchard, A. Beware of *Mycoplasma* Anti-immunoglobulin Strategies. *mBio* **2021**, *12*, e0197421. [CrossRef] [PubMed]
217. Grover, R.K.; Zhu, X.; Nieusma, T.; Jones, T.; Boreo, I.; MacLeod, A.S.; Mark, A.; Niessen, S.; Kim, H.J.; Kong, L.; et al. A structurally distinct human mycoplasma protein that generically blocks antigen-antibody union. *Science* **2014**, *343*, 656–661. [CrossRef] [PubMed]
218. Nicolson, G.L.; Nasralla, M.; Haier, J.; Pomfret, J. High frequency of systemic mycoplasmal infections in Gulf War veterans and civilians with Amyotrophic Lateral Sclerosis (ALS). *J. Clin. Neurosci.* **2002**, *9*, 525–529. [CrossRef]
219. Lo, S.C.; Levin, L.; Ribas, J.; Chung, R.; Wang, R.Y.; Wear, D.; Shih, J.W. Lack of serological evidence for *Mycoplasma fermentans* infection in army Gulf War veterans: A large scale case-control study. *Epidemiol. Infect.* **2000**, *125*, 609–616. [CrossRef] [PubMed]
220. Donta, S.T.; Engel, C.C.; Collins, J.F.; Baseman, J.B.; Dever, L.L.; Taylor, T.; Boardman, K.D.; Kazis, L.E.; Martin, S.E.; Horney, R.A.; et al. Benefits and harms of doxycycline treatment for Gulf War veterans' illnesses: A randomized, double-blind, placebo-controlled trial. *Ann. Intern. Med.* **2004**, *141*, 85–94. [CrossRef]
221. Nicolson, G.L.; Nasralla, M.; Gan, R.; Haier, J.; De Meirlier, K. Evidence for bacterial (*Mycoplasma*, *Chlamydia*) and viral (HHV-6) co-infections in chronic fatigue syndrome patients. *J. Chronic. Fatigue Syndr.* **2003**, *11*, 7–20. [CrossRef]
222. Nijs, J.; Nicolson, G.L.; De Becker, P.; Coomans, D.; De Meirlier, K. High prevalence of *Mycoplasma* infections among European chronic fatigue syndrome patients. Examination of four *Mycoplasma* species in blood of chronic fatigue syndrome patients. *FEMS Immunol. Med. Microbiol.* **2002**, *34*, 209–214. [CrossRef] [PubMed]
223. Endresen, G.K. Mycoplasma blood infection in chronic fatigue and fibromyalgia syndromes. *Rheumatol. Int.* **2003**, *23*, 211–215, Epub 2003. [CrossRef] [PubMed]
224. Carfi, A.; Bernabei, R.; Landi, F. Persistent symptoms in patients after acute COVID-19. *JAMA* **2020**, *324*, 603–605. [CrossRef] [PubMed]
225. Chu, K.A.; Ou, T.Y.; Hung, W.H.; Sung, J.; Chen, W.; Lin, C.L.; Hung, Y.M.; Wei, J.C. Mycoplasma pneumonia Infection Is Associated With an Increased Risk of Systemic Lupus Erythematosus: A Nationwide, Retrospective Cohort Study. *Front. Microbiol.* **2022**, *13*, 815136. [CrossRef] [PubMed]
226. Ginsburg, K.S.; Kundsin, R.B.; Walter, C.W.; Schur, P.H. Ureaplasma urealyticum and *Mycoplasma hominis* in women with systemic lupus erythematosus. *Arthritis Rheum.* **1992**, *35*, 429–433. [CrossRef]
227. Azizoddin, D.R.; Gandhi, N.; Weinberg, S.; Sengupta, M.; Nicassio, P.M.; Jolly, M. Fatigue in systemic lupus: The role of disease activity and its correlates. *Lupus* **2019**, *28*, 163–173. [CrossRef]
228. Iyama, K.; Zhang, S.; Lo, S.C. Effects of mycoplasmal LAMPs on receptor responses to steroid hormones in mammalian cells. *Curr. Microbiol.* **2001**, *43*, 163–169. [CrossRef]
229. Fraser, C.M.; Gocayne, J.D.; White, O.; Adams, M.D.; Clayton, R.A.; Fleischmann, R.D.; Bult, C.J.; Kerlavage, A.R.; Sutton, G.; Kelley, J.M.; et al. The minimal gene complement of *Mycoplasma genitalium*. *Science* **1995**, *270*, 397–403. [CrossRef]
230. Hutchison, C.A.; Peterson, S.N.; Gill, S.R.; Cline, R.T.; White, O.; Fraser, C.M.; Smith, H.O.; Venter, J.C. Global transposon mutagenesis and a minimal *Mycoplasma* genome. *Science* **1999**, *286*, 2165–2169. [CrossRef]
231. Almosara, J.O. Biotechnology: Genetically Engineered Pathogens. US Air Force Counterproliferation Center Future Warfare Series No. 53; 2010; Volume 53. Available online: <https://media.defense.gov/2019/Apr/11/2002115517/-1/-1/0/53ALMOSARAMONO.PDF> (accessed on 29 September 2022).
232. Sharma, A.; Gupta, G.; Ahmad, T.; Krishan, K.; Kaur, B. Next generation agents (synthetic agents): Emerging threats and challenges in detection, protection, and decontamination. *Handb. Biol. Warf. Prep.* **2020**, *217*–256. [CrossRef]
233. Busl, K.M.; Bleck, T.P. Treatment of neuroterrorism. *Neurotherapeutics* **2012**, *9*, 139–157. [CrossRef] [PubMed]
234. Malicoat, J.; Manivasagam, S.; Zuñiga, S.; Sola, I.; McCabe, D.; Rong, L.; Perlman, S.; Enjuanes, L.; Manicassamy, B. Development of a Single-Cycle Infectious SARS-CoV-2 Virus Replicon Particle System for Use in Biosafety Level 2 Laboratories. *J. Virol.* **2022**, *96*, e0183721. [CrossRef] [PubMed]
235. Margarita, V.; Fiori, P.L.; Rappelli, P. Impact of Symbiosis Between *Trichomonas vaginalis* and *Mycoplasma hominis* on Vaginal Dysbiosis: A Mini Review. *Front. Cell Infect. Microbiol.* **2020**, *10*, 179. [CrossRef] [PubMed]

236. Morris, D.E.; Cleary, D.W.; Clarke, S.C. Secondary Bacterial Infections Associated with Influenza Pandemics. *Front. Microbiol.* **2017**, *8*, 1041. [[CrossRef](#)] [[PubMed](#)]
237. Montagnier, L.; Blanchard, A. Mycoplasmas as cofactors in infection due to the human immunodeficiency virus. *Clin. Infect. Dis.* **1993**, *17* (Suppl. S1), S309–S315. [[PubMed](#)]
238. Wagner, K.D.; Wagner, N. The Senescence Markers p16INK4A, p14ARF/p19ARF, and p21 in Organ Development and Homeostasis. *Cells* **2022**, *11*, 1966. [[CrossRef](#)]
239. Kirkland, J.L.; Tchkonia, T. Senolytic drugs: From discovery to translation. *J. Intern. Med.* **2020**, *288*, 518–536. [[CrossRef](#)]
240. Juang, Y.P.; Chou, Y.T.; Lin, R.X.; Ma, H.H.; Chao, T.L.; Jan, J.T.; Chang, S.Y.; Liang, P.H. Design, synthesis and biological evaluations of niclosamide analogues against SARS-CoV-2. *Eur. J. Med. Chem.* **2022**, *235*, 114295. [[CrossRef](#)]
241. Braga, L.; Ali, H.; Secco, I.; Chiavacci, E.; Neves, G.; Goldhill, D.; Penn, R.; Jimenez-Guardeño, J.M.; Ortega-Prieto, A.M.; Bussani, R.; et al. Drugs that inhibit TMEM16 proteins block SARS-CoV-2 spike-induced syncytia. *Nature* **2021**, *594*, 88–93. [[CrossRef](#)]
242. Ansell, S.M.; Maris, M.B.; Lesokhin, A.M.; Chen, R.W.; Flinn, I.W.; Sawas, A.; Minden, M.D.; Villa, D.; Percival, M.M.; Advani, A.S.; et al. Phase I Study of the CD47 Blocker TTI-621 in Patients with Relapsed or Refractory Hematologic Malignancies. *Clin. Cancer Res.* **2021**, *27*, 2190–2199. [[CrossRef](#)]
243. Boada-Romero, E.; Martinez, J.; Heckmann, B.L.; Green, D.R. The clearance of dead cells by efferocytosis. *Nat. Rev. Mol. Cell Biol.* **2020**, *21*, 398–414. [[CrossRef](#)]
244. Heit, B.; Tasnim, T.; Le Lam, A. MER Tyrosine Kinase Mediates Efferocytosis Through a Novel  $\beta$ 2 Integrin-Activating Signalling Pathway. *J. Immunol.* **2021**, *206* (Suppl. S1), 97–104.
245. Kelley, S.M.; Ravichandran, K.S. Putting the brakes on phagocytosis: “don’t-eat-me” signaling in physiology and disease. *EMBO Rep.* **2021**, *22*, e52564. [[CrossRef](#)] [[PubMed](#)]
246. Musumeci, D.; Roviello, G.N.; Montesarchio, D. An overview on HMGB1 inhibitors as potential therapeutic agents in HMGB1-related pathologies. *Pharm. Ther.* **2014**, *141*, 347–357. [[CrossRef](#)] [[PubMed](#)]
247. Avgousti, D.C.; Herrmann, C.; Kulej, K.; Pancholi, N.J.; Sekulic, N.; Petrescu, J.; Molden, R.C.; Blumenthal, D.; Paris, A.J.; Reyes, E.D.; et al. A core viral protein binds host nucleosomes to sequester immune danger signals. *Nature* **2016**, *535*, 173–177. [[CrossRef](#)]
248. Lee, J.J.; Park, I.H.; Kwak, M.S.; Rhee, W.J.; Kim, S.H.; Shin, J.S. HMGB1 orchestrates STING-mediated senescence via TRIM30 $\alpha$  modulation in cancer cells. *Cell Death Discov.* **2021**, *7*, 28. [[CrossRef](#)]
249. Tanaka, N.; Yonekura, H.; Yamagishi, S.; Fujimori, H.; Yamamoto, Y.; Yamamoto, H. The receptor for advanced glycation end products is induced by the glycation products themselves and tumor necrosis factor-alpha through nuclear factor-kappa B, and by 17beta-estradiol through Sp-1 in human vascular endothelial cells. *J. Biol. Chem.* **2000**, *275*, 25781–25790. [[CrossRef](#)]
250. Liu, J.; Huang, K.; Cai, G.Y.; Chen, X.M.; Yang, J.R.; Lin, L.R.; Yang, J.; Huo, B.G.; Zhan, J.; He, Y.N. Receptor for advanced glycation end-products promotes premature senescence of proximal tubular epithelial cells via activation of endoplasmic reticulum stress-dependent p21 signaling. *Cell Signal.* **2014**, *26*, 110–121. [[CrossRef](#)]
251. Sourris, K.C.; Watson, A.; Jandeleit-Dahm, K. Inhibitors of Advanced Glycation End Product (AGE) Formation and Accumulation. *Handb. Exp. Pharmacol.* **2021**, *264*, 395–423. [[CrossRef](#)]
252. Zhong, H.; Li, X.; Zhou, S.; Jiang, P.; Liu, X.; Ouyang, M.; Nie, Y.; Chen, X.; Zhang, L.; Liu, Y.; et al. Interplay between RAGE and TLR4 Regulates HMGB1-Induced Inflammation by Promoting Cell Surface Expression of RAGE and TLR4. *J. Immunol.* **2020**, *205*, 767–775. [[CrossRef](#)]
253. Carlo-Stella, N.; Bozzini, S.; De Silvestri, A.; Sbarsi, I.; Pizzochero, C.; Lorusso, L.; Martinetti, M.; Cuccia, M. Molecular study of receptor for advanced glycation endproduct gene promoter and identification of specific HLA haplotypes possibly involved in chronic fatigue syndrome. *Int. J. Immunopathol. Pharmacol.* **2009**, *22*, 745–754. [[CrossRef](#)] [[PubMed](#)]
254. Hein, G.; Franke, S. Are advanced glycation end-product-modified proteins of pathogenetic importance in fibromyalgia? *Rheumatology* **2002**, *41*, 1163–1167. [[CrossRef](#)] [[PubMed](#)]
255. Nicotra, L.; Loram, L.C.; Watkins, L.R.; Hutchinson, M.R. Toll-like receptors in chronic pain. *Exp. Neurol.* **2012**, *234*, 316–329. [[CrossRef](#)] [[PubMed](#)]
256. Alhasson, F.; Das, S.; Seth, R.; Dattaroy, D.; Chandrashekaran, V.; Ryan, C.N.; Chan, L.S.; Testerman, T.; Burch, J.; Hofseth, L.J.; et al. Altered gut microbiome in a mouse model of Gulf War Illness causes neuroinflammation and intestinal injury via leaky gut and TLR4 activation. *PLoS ONE* **2017**, *12*, e0172914. [[CrossRef](#)]
257. Fontes-Dantas, F.L.; Fernandes, G.G.; Gutman, E.G.; De Lima, E.V.; Antonio, L.S.; Hammerle, M.B.; Mota-Araujo, H.P.; Colodeti, L.C.; Araújo, S.M.B.; da Silva, T.N.; et al. SARS-CoV-2 spike protein induces long-term TLR4-mediated synapse and cognitive loss recapitulating Post-COVID syndrome. *bioRxiv* **2022**. [[CrossRef](#)]
258. Zhang, Y.; Liang, X.; Bao, X.; Xiao, W.; Chen, G. Toll-like receptor 4 (TLR4) inhibitors: Current research and prospective. *Eur. J. Med. Chem.* **2022**, *235*, 114291. [[CrossRef](#)]
259. Snelson, M.; Lucut, E.; Coughlan, M.T. The Role of AGE-RAGE Signalling as a Modulator of Gut Permeability in Diabetes. *Int. J. Mol. Sci.* **2022**, *23*, 1766. [[CrossRef](#)]
260. Paudel, Y.N.; Angelopoulou, E.; Semple, B.; Piperi, C.; Othman, I.; Shaikh, M.F. Potential Neuroprotective Effect of the HMGB1 Inhibitor Glycyrhizin in Neurological Disorders. *ACS Chem. Neurosci.* **2020**, *11*, 485–500. [[CrossRef](#)]
261. Oh, S.H.; Lee, H.Y.; Ki, Y.J.; Kim, S.H.; Lim, K.J.; Jung, K.T. Gabexate mesilate ameliorates the neuropathic pain in a rat model by inhibition of proinflammatory cytokines and nitric oxide pathway via suppression of nuclear factor- $\kappa$ B. *Korean J. Pain.* **2020**, *33*, 30–39. [[CrossRef](#)]

262. Gobbiotti, T.; Cenac, N.; Motta, J.P.; Rolland, C.; Martin, L.; Andrade-Gordon, P.; Steinhoff, M.; Barocelli, E.; Vergnolle, N. Serine protease inhibition reduces post-ischemic granulocyte recruitment in mouse intestine. *Am. J. Pathol.* **2012**, *180*, 141–152. [CrossRef]
263. Nishibori, M.; Mori, S.; Takahashi, H.K. Anti-HMGB1 monoclonal antibody therapy for a wide range of CNS and PNS diseases. *J. Pharmacol. Sci.* **2019**, *140*, 94–101. [CrossRef] [PubMed]
264. Sgrignani, J.; Cecchinato, V.; Fassi, E.M.A.; D’Agostino, G.; Garofalo, M.; Danelon, G.; Pedotti, M.; Simonelli, L.; Varani, L.; Grazioso, G.; et al. Systematic Development of Peptide Inhibitors Targeting the CXCL12/HMGB1 Interaction. *J. Med. Chem.* **2021**, *64*, 13439–13450. [CrossRef] [PubMed]
265. Ishibashi, Y.; Matsui, T.; Isami, F.; Abe, Y.; Sakaguchi, T.; Higashimoto, Y.; Yamagishi, S.I. N-butanol extracts of Morinda citrifolia suppress advanced glycation end products (AGE)-induced inflammatory reactions in endothelial cells through its anti-oxidative properties. *BMC Complement. Altern. Med.* **2017**, *17*, 137. [CrossRef] [PubMed]
266. Batista, J.A.; Magalhães, D.A.; Sousa, S.G.; Ferreira, J.D.S.; Pereira, C.M.C.; Lima, J.; de Albuquerque, I.F.; Bezerra, N.L.S.D.; de Brito, T.V.; Monteiro, C.E.D.S.; et al. Polysaccharides derived from Morinda citrifolia Linn reduce inflammatory markers during experimental colitis. *J. Ethnopharmacol.* **2020**, *248*, 112303. [CrossRef] [PubMed]
267. Li, X.; Liu, Y.; Shan, Y.; Wang, Y.; Li, Z.; Bi, Y.; Zhao, W.; Yin, Y.; Wang, T.; Li, S.; et al. MicroRNAs Involved in the Therapeutic Functions of Noni (*Morinda citrifolia* L.) Fruit Juice in the Treatment of Acute Gouty Arthritis in Mice Induced with Monosodium Urate. *Foods* **2021**, *10*, 1638. [CrossRef]
268. Lucas, K.; Maes, M. Role of the Toll Like receptor (TLR) radical cycle in chronic inflammation: Possible treatments targeting the TLR4 pathway. *Mol. Neurobiol.* **2013**, *48*, 190–204. [CrossRef]
269. Su, K.; Bo, L.; Jiang, C.; Deng, X.; Zhao, Y.Y.; Minshall, R.D.; Hu, G. TLR4 is required for macrophage efferocytosis during resolution of ventilator-induced lung injury. *Am. J. Physiol. Lung Cell. Mol. Physiol.* **2021**, *321*, L787–L801. [CrossRef]
270. Liu, Y.; Li, J.; Lu, X.; Zhen, S.; Huo, J. Toll-Like Receptor 4 Exacerbates Mycoplasma pneumoniae Promoting Transcription Factor EB-Mediated Autophagy. *Contrast Media Mol. Imaging* **2022**, *2022*, 3357694. [CrossRef]
271. Anwar, M.A.; Panneerselvam, S.; Shah, M.; Choi, S. Insights into the species-specific TLR4 signaling mechanism in response to Rhodobacter sphaeroides lipid A detection. *Sci. Rep.* **2015**, *5*, 7657. [CrossRef]
272. Tam, J.S.Y.; Coller, J.K.; Hughes, P.A.; Prestidge, C.A.; Bowen, J.M. Toll-like receptor 4 (TLR4) antagonists as potential therapeutics for intestinal inflammation. *Indian J. Gastroenterol.* **2021**, *40*, 5–21. [CrossRef]
273. Ono, Y.; Maejima, Y.; Saito, M.; Sakamoto, K.; Horita, S.; Shimomura, K.; Inoue, S.; Kotani, J. TAK-242, a specific inhibitor of Toll-like receptor 4 signalling, prevents endotoxemia-induced skeletal muscle wasting in mice. *Sci. Rep.* **2020**, *10*, 694. [CrossRef] [PubMed]
274. Masaldan, S.; Clatworthy, S.A.S.; Gamell, C.; Meggyesy, P.M.; Rigopoulos, A.T.; Haupt, S.; Haupt, Y.; Denoyer, D.; Adlard, P.A.; Bush, A.I.; et al. Iron accumulation in senescent cells is coupled with impaired ferritinophagy and inhibition of ferroptosis. *Redox Biol.* **2018**, *14*, 100–115. [CrossRef] [PubMed]
275. Chen, Y.; Xu, Y.; Zhang, K.; Shen, L.; Deng, M. Ferroptosis in COVID-19-related liver injury: A potential mechanism and therapeutic target. *Front. Cell Infect. Microbiol.* **2022**, *12*, 922511. [CrossRef] [PubMed]
276. Sfera, A.; Bullock, K.; Price, A.; Inderias, L.; Osorio, C. Ferrosenescence: The iron age of neurodegeneration? *Mech. Ageing* **2018**, *174*, 63–75. [CrossRef]
277. Binder, C.J.; Papac-Milicevic, N.; Witztum, J.L. Innate sensing of oxidation-specific epitopes in health and disease. *Nat. Rev. Immunol.* **2016**, *16*, 485–497. [CrossRef]
278. Leibundgut, G.; Witztum, J.L.; Tsimikas, S. Oxidation-specific epitopes and immunological responses: Translational biotheranostic implications for atherosclerosis. *Curr. Opin. Pharmacol.* **2013**, *13*, 168–179. [CrossRef]
279. Osthues, T.; Sisignano, M. Oxidized Lipids in Persistent Pain States. *Front. Pharmacol.* **2019**, *10*, 1147. [CrossRef]
280. Liang, H.; Luo, D.; Liao, H.; Li, S. Coronavirus Usurps the Autophagy-Lysosome Pathway and Induces Membranes Rearrangement for Infection and Pathogenesis. *Front. Microbiol.* **2022**, *13*, 846543. [CrossRef]
281. Wang, S.; Li, W.; Zhang, P.; Wang, Z.; Ma, X.; Liu, C.; Vasilev, K.; Zhang, L.; Zhou, X.; Liu, L.; et al. Mechanical overloading induces GPX4-regulated chondrocyte ferroptosis in osteoarthritis via Piezo1 channel facilitated calcium influx. *J. Adv. Res.* **2022**; *in press*. [CrossRef]
282. Pedrera, L.; Espiritu, R.A.; Ros, U.; Weber, J.; Schmitt, A.; Stroh, J.; Hailfinger, S.; von Karstedt, S.; García-Sáez, A.J. Ferroptotic pores induce Ca<sup>2+</sup> fluxes and ESCRT-III activation to modulate cell death kinetics. *Cell Death Differ.* **2021**, *28*, 1644–1657. [CrossRef]
283. Ayala, A.; Muñoz, M.F.; Argüelles, S. Lipid Peroxidation: Production, Metabolism, and Signaling Mechanisms of Malondialdehyde and 4-Hydroxy-2-Nonenal. *Oxid. Med. Cell. Longev.* **2014**, *2014*, 360438. [CrossRef]
284. Maes, M.; Kubera, M.; Uytterhoeven, M.; Vrydags, N.; Bosmans, E. Increased plasma peroxides as a marker of oxidative stress in myalgic encephalomyelitis/chronic fatigue syndrome (ME/CFS). *Med. Sci. Monit.* **2011**, *17*, SC11–SC15. [CrossRef] [PubMed]
285. Shukla, V.; Kumar, D.S.; Ali, M.A.; Agarwal, S.; Khandpur, S. Nitric oxide, lipid peroxidation products, and antioxidants in primary fibromyalgia and correlation with disease severity. *J. Med. Biochem.* **2020**, *39*, 165–170. [CrossRef] [PubMed]
286. Naviaux, R.K.; Naviaux, J.C.; Li, K.; Wang, L.; Monk, J.M.; Bright, A.T.; Koslik, H.J.; Ritchie, J.B.; Golomb, B.A. Metabolic features of Gulf War illness. *PLoS ONE* **2019**, *14*, e0219531. [CrossRef] [PubMed]
287. Zhang, X.; Song, T.; Zhao, M.; Tao, X.; Zhang, B.; Sun, C.; Wang, P.; Wang, K.; Zhao, L. Sirtuin 2 Alleviates Chronic Neuropathic Pain by Suppressing Ferroptosis in Rats. *Front. Pharmacol.* **2022**, *13*, 827016. [CrossRef] [PubMed]

288. Reichert, C.O.; de Freitas, F.A.; Sampaio-Silva, J.; Rokita-Rosa, L.; Barros, P.L.; Levy, D.; Bydlowski, S.P. Ferroptosis Mechanisms Involved in Neurodegenerative Diseases. *Int. J. Mol. Sci.* **2020**, *21*, 8765. [[CrossRef](#)]
289. Wen, Q.; Liu, J.; Kang, R.; Zhou, B.; Tang, D. The release and activity of HMGB1 in ferroptosis. *Biochem. Biophys. Res. Commun.* **2019**, *510*, 278–283. [[CrossRef](#)]
290. Li, S.; Zhou, C.; Zhu, Y.; Chao, Z.; Sheng, Z.; Zhang, Y.; Zhao, Y. Ferrostatin-1 alleviates angiotensin II (Ang II)- induced inflammation and ferroptosis in astrocytes. *Int. Immunopharmacol.* **2021**, *90*, 107179. [[CrossRef](#)]
291. von Mässenhausen, A.; Zamora Gonzalez, N.; Maremonti, F.; Belavgeni, A.; Tonnus, W.; Meyer, C.; Beer, K.; Hannani, M.T.; Lau, A.; Peitzsch, M.; et al. Dexamethasone sensitizes to ferroptosis by glucocorticoid receptor-induced dipeptidase-1 expression and glutathione depletion. *Sci. Adv.* **2022**, *8*, eabl8920. [[CrossRef](#)]
292. Nicolson, G.L.; Ferreira de Mattos, G.; Ash, M.; Settineri, R.; Escribá, P.V. Fundamentals of Membrane Lipid Replacement: A Natural Medicine Approach to Repairing Cellular Membranes and Reducing Fatigue, Pain, and Other Symptoms While Restoring Function in Chronic Illnesses and Aging. *Membranes* **2021**, *11*, 944. [[CrossRef](#)]
293. Nicolson, G.L. Membrane Lipid Replacement—A functional approach to repairing cellular membranes, reducing symptoms, and restoring function. *Funct. Food Sci.* **2022**, *2*, 198–204.
294. Marchetti, G.; Tincati, C.; Silvestri, G. Microbial translocation in the pathogenesis of HIV infection and AIDS. *Clin. Microbiol. Rev.* **2013**, *26*, 2–18. [[CrossRef](#)] [[PubMed](#)]
295. Pinching, A.J. AIDS and CFS/ME: A tale of two syndromes. *Clin. Med.* **2003**, *3*, 78–82. [[CrossRef](#)] [[PubMed](#)]
296. Giron, L.B.; Peluso, M.J.; Ding, J.; Kenny, G.; Zilberstein, N.F.; Koshy, J.; Hong, K.Y.; Rasmussen, H.; Miller, G.; Bishehsari, F.; et al. Markers of Fungal Translocation Are Elevated During Post-Acute Sequelae of SARS-CoV-2 Infection and Induce NF- $\kappa$ B Triggered Inflammation. *bioRxiv* **2022**. [[CrossRef](#)]
297. Michailidis, C.; Giannopoulos, G.; Vigklis, V.; Armenis, K.; Tsakris, A.; Gargalianos, P. Impaired phagocytosis among patients infected by the human immunodeficiency virus: Implication for a role of highly active anti-retroviral therapy. *Clin. Exp. Immunol.* **2012**, *167*, 499–504. [[CrossRef](#)] [[PubMed](#)]
298. Silverstein, N.J.; Wang, Y.; Manickas-Hill, Z.; Carbone, C.; Dauphin, A.; Boribong, B.P.; Loiselle, M.; Davis, J.; Leonard, M.M.; Kuri-Cervantes, L. Innate lymphoid cells and COVID-19 severity in SARS-CoV-2 infection. *Elife* **2022**, *11*, e74681. [[CrossRef](#)] [[PubMed](#)]
299. Kløverpris, H.N.; Kazer, S.W.; Mjösberg, J.; Mabuka, J.M.; Wellmann, A.; Ndhlovu, Z.; Yadon, M.C.; Nhamoyebonde, S.; Muenchhoff, M.; Simoni, Y.; et al. Innate Lymphoid Cells Are Depleted Irreversibly during Acute HIV-1 Infection in the Absence of Viral Suppression. *Immunity* **2016**, *44*, 391–405. [[CrossRef](#)]
300. Kong, X.; Feng, D.; Wang, H.; Hong, F.; Bertola, A.; Wang, F.S.; Gao, B. Interleukin-22 induces hepatic stellate cell senescence and restricts liver fibrosis in mice. *Hepatology* **2012**, *56*, 1150–1159. [[CrossRef](#)]
301. Yang, T.; Yang, Y.; Wang, D.; Li, C.; Qu, Y.; Guo, J.; Shi, T.; Bo, W.; Sun, Z.; Asakawa, T. The clinical value of cytokines in chronic fatigue syndrome. *J. Transl. Med.* **2019**, *17*, 213. [[CrossRef](#)]
302. Wei, H.X.; Wang, B.; Li, B. IL-10 and IL-22 in Mucosal Immunity: Driving Protection and Pathology. *Front. Immunol.* **2020**, *11*, 1315. [[CrossRef](#)]
303. Vogl, T.; Kalka, I.N.; Klompus, S.; Leviatan, S.; Weinberger, A.; Segal, E. Systemic antibody responses against human microbiota flagellins are overrepresented in chronic fatigue syndrome patients. *Sci. Adv.* **2022**, *8*, eabq2422. [[CrossRef](#)]
304. König, R.S.; Albrich, W.C.; Kahlert, C.R.; Bahr, L.S.; Löber, U.; Vernazza, P.; Scheibenbogen, C.; Forslund, S.K. The Gut Microbiome in Myalgic Encephalomyelitis (ME)/Chronic Fatigue Syndrome (CFS). *Front. Immunol.* **2022**, *12*, 628741. [[CrossRef](#)] [[PubMed](#)]
305. Jahanbani, F.; Maynard, R.D.; Sing, J.C.; Jahanbani, S.; Perrino, J.J.; Spacek, D.V.; Davis, R.W.; Snyder, M.P. Phenotypic characteristics of peripheral immune cells of Myalgic encephalomyelitis/chronic fatigue syndrome via transmission electron microscopy: A pilot study. *PLoS ONE* **2022**, *17*, e0272703. [[CrossRef](#)] [[PubMed](#)]